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Significances and importance of phytochemical present in Terminalia chebula

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Abstract: Phytochemical are naturally present in the plants and shows biologically significance by playing an essential role in the plants to defend themselves against various pathogenic microbes by showing the antimicrobial activity by inhibition or killing mechanisms. The secretion of these compounds is varying from plant to plant some produce more and some produce in minimal quantity. Sometimes they can be harmful and sometimes they can be very helpful. There is evidence from laboratory studies that phytochemicals in fruits and vegetables may reduce the risk of cancer, possibly due fibers, polyphenol antioxidants and anti-inflammatory effects. phytochemicals, such as fermentable dietary fibers, are allowed limited health claims by the US Food and Drug Administration The present study was focused to find out the photochemical analysis of Terminalia chebula plant extracts of leaves, fruits, seed, stem and roots. The formation of yellow colour indicated the presence of flavonoids while the brown colour formation indicated the presence alkaloids and terpenoids. The phenol content was maximum in roots (82.13 mg/gdw) followed by seed leave, stem and fruit. The sugar content was highest in leaves (8.27 mg/gdw) followed by fruits, stem, root and seed. The protein content was maximum in fruits (55.59 mg/dgw) followed by seeds leaves, stem and root.

Keywords: phenol, terpenoids, flavonoids, alkaloid, protein, secondary metabolites

ntroduction

Medicinal plants are rich source of novel drugs that forms the ingredients in traditional systems of medicine, modern medicines, nutraceuticals, food folk medicines, supplements, intermediates bioactive pharmaceuticals principles and lead compounds in synthetic drugs [1]. One of the most important medicinal plants, which are widely used in the traditional system of medicine, is Terminalia chebula [2]. This drives the need to screen medicinal plants for novel bioactive compounds as plant based drugs are biodegradable and safe [3]. A natural product plays an important role in the field of new drugs research and development because of their low toxicity, easy availability and cost effective [4]. The primary metabolite like chlorophyll, amino

acids, nucleotides, simple carbohydrates or membrane lipids, play recognized roles in photosynthesis, respiration, solute transport, translocation. nutrient assimilation and differentiation [5]. Secondary metabolites are synthesized by the plants as part of the defense system of the plant [6]. The plant contains chebulinc acid, tannic acid, gallic acid, resin, anthroquinone and sennoside. It also contains glycosides, sugar, triterpenoids, steroids and small quantity of phosphoric acid these compounds were proven to exhibit anti-bacterial, anti-fungal, anti-viral and anti-carcinogenic [7]. The Terminalia chebula shows antioxidant, adaptogenic and anti-anaphylactic, hypolipidemic, hepato protective, anti-diabetic, protective, cardio wound healing, immuno-modulatory and chemo preventive [8, 9]. Terminalia chebula is rich in tannin, which is hydrolysable to pyrogallal was found in fruits [10].

Materials & Methods

Collection of Plant

Herbal Plant *Terminalia chebula* was collected from the dense forest at Bengali, Karnataka state, India, based on its ethno medical importance.

Preparation of powder

The leaves, fruits, seed, stem and root powders were prepared [11] by washing with distilled water, surface sterilized with 10% sodium hypochlorite solution, rinsed with sterile distilled water and air dried at room temperature under shadow and then milled to fine powder.

Extraction of plant material

Methanolic extract

The methanol extract was prepared [4] by taking 10 grams of powdered sample, were soaked in 50 ml of methanol and it was kept in Soxhled apparatus at 80 degree Celsius for 48 hours. This extraction was taken and allowed for evaporation and it was concentrated with Dimethyl Sulfoxide (4.64g).

Phytochemical activity

The phytochemical analysis of Terminalia chebula was carried out [12] by adopting the following tests

Test for alkaloids

The Terminalia chebula extracts of leaves, fruits, seed, stem and root were filtrated then treated with Potassium mercuric iodide (Dragendroffs reagent) and observed for the colour change in the test tubes.

Test for flavonoids

0.2 grams of plant extracts such as leaves, fruits, stem and root was added into test tube

containing 2ml of diluted sodium hydroxide and mixed well. After mixing 2ml of diluted hydrochloride was added into the test tubes and observed for colour change.

Test for terpenoids

0.5grams of plant extracts such as leaves, fruits, seed, stem and root was added to test tubes containing 2ml of chloroform and content was mixed well. Then 2ml of concentrated sulphuric acid was added carefully and observe for presence of reddish brown colour.

Quantities assay for tannin

The quantities assay for tannin acids was carried [6] out by taking the 500mg finely dried plant extract was added into a glass beaker containing 5ml of 70% aqueous acetone. The content solution was uniformly mixed and gently boiled in a water bath for 30 minutes. The solution was centrifuged at 3000 rpm for 10 minutes at 4°C and supernatants were collected and stored in freezing condition. The pallet were dissolved in 5ml of 70% aqueous acetone and recentrifuged at 3000rpm for 10 minutes at 4°C. The supernatants were collected and mixed with freezing stored supernatants. To this supernatants 1 ml of Folindenis reagent, 3ml of Sodium carbonate solution was added and solution was diluted to 20 ml by using distilled water. The solution was mixed well and incubated at room temperature for 30 minutes. The absorbance was measured in a spectrophotometer at 700nm.

Extraction of Reducing sugar

Extraction of sugars from the sample was usually carried [9] out by using 80 per cent ethanol. The crushed material is refluxed in a Soxhlet's apparatus for about 30 minutes. After refluxing, alcohol was wiped out from the extract with the addition of distilled water. The extract was centrifuged at 3000 rpm .To the supernatant, 1 ml

of saturated lead acetate solution and 1 ml of saturated Di-sodium hydrogen phosphate solution was added. After centrifugation, the clear supernatant was collected and made up to a known volume with distilled water. To make standard, 1ml aliquots are taken from the supernatant, respectively in hard-glass test tubes. The 5ml of Somogyi's reagent and distilled water are added so that final volume of the solution in each test tube comes to 15ml. The tubes covered with lids are heated in a boiling water bath for about 15 minutes. These are then cooled to room temperature. The tubes, in which the solutions have reprecipitated, are discarded. 1ml of 2.5 per cent potassium iodide solution was added to each test tube. Then 3ml of 1.5N Sulphuric acid was added to this and shaken well till the golden yellow colour is formed. A burette was filled with 0.005 N sodium thiosulphate solutions. It was titrated with the sample on addition of 1-2 drops of starch solution to the latter. The end point was determined by the complete disappearance of the colour from the solution. A blank was also prepared by mixing 5ml of Somogyi's reagent and 10ml of distilled water. It was heated in a boiling water bath for about 15 minutes. After cooled and 1 ml of 2.5 per cent potassium iodide solution and 3ml of 1.5 N sulphuric acid are added. Titration was done using 0.001 N sodium thiosulphate solution containing starch solution and colorless end-point was determined.

Identification of amino acid, phenolic and aromatic compounds by using TLC method

A sample of 500mg/ml concentration of plant extracts was prepared [13] and from this solution, 4µl of the sample prepared was taken and spotted on the silica coated TLC plate. It was then kept at solvent position with the solvent to run under capillary pressure. Here ethanol, methanol and acetic acid (5ml, 5ml, and 100µl) were used as a solvent. The spots were then identified in the iodine chamber for phenolic compounds, ferric chloride for aromatic compounds and ninhydrin for amino acids.

Column chromatography

The crude aqueous methanol extract Terminalia chebula was subjected to column chromatography [1] over silica gel 100-200 mesh. The column was eluted with solvents of increasing order of polarity. The fractions were collected in 10ml each and allowed to evaporate to get the residue. Each fraction was tested for the presence various constituents by Thin Layer Chromatography.

RESULT

Terminalia chebula

Kingdom-Plantae, Subkingdom-Tracheobionta, Superdivision-Spermatophyta, Magnoliophyta, Class-Magnoliopsida, Subclass-Rosidae, Order-Myrtales, Family-Combretaceae, Genus-Terminalia, Species-Terminalia chebula

Phytochemical activity:

Alkaloid

There was a reddish brown colour formation in the test tubes after treating with Potassium Bismuth iodide solution thus indicates the presence of alkaloid in the plant extract (Figure 1).

Flavonids

The yellow colour was formed in the test tubes after treating with 5% Sodium hydroxide and diluted hydrochloride acid thus indicated the presence of flavonoids (Figure 2).

Terpenoid

The Terpenoid was present in the plant extract as there was reddish brown colour formation in the The phenol content was found maximum in root (85.36mg/gdw) followed by seed (78.30mg/gdw), stem (72.46mg/gdw), roots (65.30mg/gdw) and leaves (21.39mg/gdw).

Reducing sugar

The maximum content of sugar was in leaves (7.12mg/gdw) followed by fruit (5.70mg/gdw), stem (4.80mg/gdw), root (4.10mg/gdw) and seed (3.7mg/gdw).

Thin layer Chromatography

In thin layer chromatography the amino acids showed pink colour spots were observed after

treating with ninhydrin. The sugars showed the purple and black spots after treating with ferric chloride. The phenolic compounds showed blue spots after treating with iodine solution

Column chromatography

Higher content of protein was observed in fruits of T. Chebula (44.40mg/gdw) followed by seed (42.10mg/gdw), roots (40.60mg/gdw), leaves (36.10mg/gdw) and stem (29.40mg/gdw). The Purification of compounds in various extracts was performed by using column chromatography was presented in (Table 1).

Table 1: Purification of methanol extracts compounds of *Terminalia chebula* by using column chromatography

No of fractions	Eluent	Ratio	Colours of fraction	Rf value
F1-F2	Acetone	100	Brown	1.52
F3-F5	Acetone, Ethanol	75:25, 50:50, 25:75	Dark Brown	1.6
F6-F7	Acetone, Ethanol	75:25, 50:50, 25:75	Dark Brown	1.25, 1.18
F8-F13	Ethanol	100	Light Brown	1.51
F14-F18	Ethanol, Methanol	75:25, 50:50, 25:75	Colourless	-
F19-F40	Methanol	100	Colourless	-

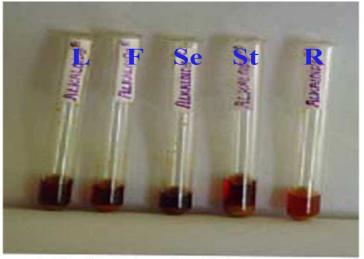


Figure 1. The formation of reddish brown colour in the plant extracts of *Terminalia chebula* when treated with Potassium Bismuth solution indicated the presence of Alkaloids. Note where L--Leaf, F--Fruit, Se--Seed, St--Stem, R--Root

Figure 2. The formation of yellow colour in the plant extracts of Terminalia chebula when treated with 5% Sodium Hydroxide solution containing diluted Hydrochloride solution thus indicated the presenc of Flavoniods. Note where L--Leaf, F--Fruit, Se--Seed, St-Seed.



Figure 3. The formation of redish brown colour in the plant extracts of Terminalia chebula when treated with chloroform containing concertrated Sulphuric acid solutin thus indicated the presence of Terpenoids. Note where L--Leaf, F--Fruit, Se--Seed, St--Stem,

DISCUSION

Terminalia chebula is called the king of medicine in Tibet and is always listed at the top of the listed in Ayurvedic medicine [14]. A diverse arsenal of new antibacterial agents is urgently needed to combat the diminishing efficacy of existing antibiotics [15]. In earlier studies, lipids were reported to be higher in leaves (46.0mg/gdw) of M. oleifera and the roots (39.0mg/gdw) of C. quadrangularis [16, 17]. In modern therapeutic treatments, nanoparticles and liposomes are being used to develop delivery systems that are convenient and effective for tackling problems in disease treatments [18]. Proteins are complex nitrogenous organic substances that are one of the most important plant products to man. A part from this, the protein hydrolytes from various sources are reported to possess antioxidant activity [19, 20]. Plant phenols are groups of natural products with variable structure that are well known for their beneficial effects on health possess significant antimicrobial and antioxidant activities [21, 22]. Terminalia chebula have been noted to possess shikimic acid, gallic acid, B sitosterol, tannic acid, triethyl ester of chebulic acid, ethylester of gallic acid and ellagic ethaedioic acid [23]. Terminalia plant contains

several constituents like tannins, flavonoids, sterols, amino acids, fructose, resin, and fixed oils. It is also found to contain compounds like anthraquinones, 4, 2, 4 chebulyl-dglucopyranose, terpinenes and terpinenols [24]. metabolite also acts as precursors for bioactive compounds used as therapeutic drugs [25]. Phytochemical from medicinal plants serve as lead compounds in drug discovery and design Aellagitann interchebulin [26]. along with punicalagin, terflavin-A, shikimic, tricontanoic, palmitic acids, beta-sitosterol, daucosterol, triterpene chebupentol were found in fruits [27]. The compounds phloroglucinol and pyrogallol, isolated along with ferulic, vanillic, p-coumaric and caffeic acids constitutes for the antioxidant activity of the plant [28].

References

- Praveen N, Nayak S, Kar DM and Das P. Pharmacological evaluation of ethanolic extracts of the plant Alternanthera sessilis against temperature regulation. J. Pharm. Res., 2010; 3(6):1381-1383.
- Anitha M, Satish Kumar BN, Vrushabendra Swamy BM and Archana S. A Review on Natural Diuretics. Res. J. of Pharm. Biolo. and Chem. Scien.2010;1(4):615-634.
- Ramakrishna S, Ramana KV, Mihira V and Kumar BP. Evaluation of anti-inflammatory and analgesic activities of Solanum trilobatum Linn. Roots. Res. J. Pharm. Biol. Chem. Sci.200; 2(1):701-705.
- Chessbrough M. Medicinal laboratory manual for tropical countries, Lianacre Houses. Volume II: Microbiology. Chap. 200;44:289-311. Butterworth-Heinemann Ltd., Linacre House, Jordan, Hill, Oxford.
- 5) Taiz Q and Zeiger E. Plant physiology 4th Edn. 2006:13: PP 315-344.

- 6) Phan TT, Wang L, See P, Grayer RJ, Chan SY and Lee ST. Phenolic Compounds of Chromolaena odorata Protect Cultured Skin Cells from Oxidative Damage: Implication for Cutaneous Wound Healing. Biol. Pharm. Bull.2001; 24(12):1373-1379.
- 7) Neamsuvan O, Singdam P, Yingcharoen K and Sengnon N (2012). A survey of medicinal plants in mangrove and beach forests from sating Phra Peninsula, Songkhla Province, Thailand. Journal of Med. Plants Research. 6(12):2421-2437.
- 8) Chattopadhyay RR, Bhattacharyya SK, Anwesa B, Premananda B, and Nitish Kumar P. Evaluation of antibacterial properties of Chebulic myrobalan (fruit of Terminalia chebula Retz.) extracts against methicillin resistant Staphylococcus aureus and trimethoprimsulphamethoxazole resistant uropathogenic Escherichia coli. African J. of Plant Scien. 2009; 3(2): 025-029.
- 9) Souza LD, Wahidulla S, and Devi P. Antibacterial phenolics from the mangrove *Lumnitzera* racemosa. Indian J. Mar. Sci.2010; 39(2):294-298.

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- 10) Ghosh A, Das BK, Roy A, Mandal B, and Chanda G. Antimicrobial activity of some medicinal plants extracts. Journal of Nat. Med. 2008; 62(2):259-262.
- 11) Ncube NS, Afolayan AJ and Okah Al.
 Assessment techniques of antimicrobial properties of natural compounds of plant origin:current methods and future trends. African Journal of Biotechnology. 2008; 7 (12):1797-1806.
- 12) Yankanchi SR and Koli SA. Anti-inflammatory and Analgesic activity of mature leaves methanol extract of *Clerodendrum inerme* L. (Gaertn). J. Pharm. Sci. Res. 2010; 2(11):782-785.]
- 13) Kiyota H. Synthesis of naturally derived bioactive compounds of agricultural interest. Biosci Biotechnol Biochem. 2006; 70(2):317-24.
- 14) Kannan P, Ramadevi SA and Waheeta H. Antibacterial activity of *Terminalia chebula* fruit extract. Journal of Microbiology. 2009; 3(4):180-184.

- 15) Ahmad I, Mehmood Z and Mohammad F. Screening of some Indian medical plant for their antimicrobial properties. J. Ethnopharmacon. 1989; 62(2):183-193.
- 16) Bussmann RW, Gilbreath GG, Solio J, Lutura M, Lutuluo R, Kunkuru K, Wood N and Mathenge SG. Plant use of the Maasai of Sekenani Valley, Maasai Mara, Kenya. J. Ethnobiol. Ethnomed., 2006;2: 22, doi: 10.1186/1746-4269-2-22
- 17) Shariff N, Sudarshana MS, Umesha S and Hariprasad P. Antimicrobial activity of *Rauvolfia tetraphylla* and *Physalis minima* leaf and callus extracts. Afr. J. Biotechnol. 2006; 5(10):946-950.
- 18) Tariq, AL and Reyaz AL. Therapeutic role of Terminalia Chebula against Uropathogenic Escherichia Coli (UPEC). Global Journal of Pharmacology. 2012; 6 (3): 160-165.
- 19) Luziatelli G, Sorensen M, Theilade I and Molgaard P. Ashaninka medicinal plants: a case study from the native community of Bajo Quimiriki, Junín, Peru. J. Ethnobiol. Ethnomed.2010; 6:21-27
- 20) Shah BN, Nayak BS, Seth AK, Jalalpure SS, Patel KN, Patel MA and Mishra AD. Review Article Search for medicinal plants as a source of anti-inflammatory and anti-arthritic agents. Pharmacogn. Mag., 2006;2(6):77-86.
- 21) Prashith KTR, Vinayaka KS, Soumya KV, Ashwini SK and Kiran R. Antibacterial and antifungal activity of methanolic extract of *Abrus pulchellus* wall and *Abrus precatorius* Linn- A comparative study. Int. J. Toxicol. Pharmacol. Res. 2010; 2(1):26-29.
- 22) Sahu NP and Mahato SB. Anti-inflammatory triterpene saponins of Pithecellobium dulce: characterization of an echinocystic acid bisdesmoside. Phytochemistry. 1994; 37(5):1425-1427.
- 23) Ates DA and Erdourul OT. Antimicrobial Activities of Various Medicinal and Commercial Plant Extracts. Truk. Journal of Biol. 2003; 27:157-162.
- 24) Archana S, Abhishek C, Madhuliki S, Farrukh J, Preethi P, Siron MR, Falgun WB and Lakshmi V.

- Inhibition of Hyaluronidase activity of human and rat spermatozoa invitro by *Terminalia*, Flavanoid richplant. Reproductive Toxicology. 2010; 29 (2):214-224.
- 25) Ebi GC and Ofoefule SI. Antimicrobial activity of Pterocarpus osun stems. Fitoterapia. 2000; 71(4):433-435.
- 26) Falodun A and Agbakwuru EOP. Phytochemical analysis and laxative activity of the leaf extracts of Euphobia heterophylla Linn. (Euphorbiaceace). Pak. J. Sci. Ind. Res., 2004; 47(5): 345-348.
- 27) Luziatelli G, Sorensen M, Theilade I and M0lgaard P. Ashaninka medicinal plants: a case study from the native community of Bajo Quimiriki, Junín, Peru. J. Ethnobiol. Ethnomed., 2010;6: 21.
- 28) Mazumder UK, Gupta M, Manikandan L, Bhattacharya S, Haldar PK and Roy S. Evaluation of anti-inflammatory activity of Vernonia cinerea Less. extract in rats. Phytomedicine, 2003; 10:185-188.

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