

International Journal of Drug Development & Research | July-Sept 2010 | Vol. 2 | Issue 3 | ISSN 0975-9344 | Available online http://www.ijddr.in Covered in Official Product of Elsevier, The Netherlands ©2010 IJDDR

PREPARATION AND EVALUATION OF SODIUM DICLOFENAC LOADED CHITOSAN CONTROLLED RELEASE MICROPARTICLES USING FACTORIAL DESIGN

Behzad Sharif Makhmal Zadeha, Fatemeh Moshtaghi b, Fakher Rahime, Abbas akhgarib

^aNanotechnology Research Center, Ahvaz Jundishapur University of Medical Sciences ^bSchool of pharmacy, Ahvaz Jundishapur University of Medical Sciences ^cPhysiology Research Center, Ahvaz Jundishapur University of medical sciences

ABSTRACT

Sodium diclofenac is a non-steroidal anti-inflammatory drug used to relieve the inflammation, swelling, stiffness, and joint pain associated with rheumatoid arthritis. Chitosan is a linear polysaccharide derived from chitin that has been used in different pharmaceutical forms and drug delivery systems. The purpose of this study was to prepare and evaluate the chitosan beads as a controlled drug release system for sodium diclofenac consists of sustained release microparticles and immediate released particles. Sustained release chitosan microparticles were prepared with an ionic cross-linking agent. Loading efficiency was influenced by percentage of sodium polyphosphate, Tween and drug-chitosan proportion with the maximum amount of 83.12%. Release of sodium diclofenac from microparticles pattern indicated burst and sustained release that was influenced by percentage of Tween 80. Finally, Chitosan microparticles provided suitable sustained release and drug loading for sodium diclofenac oral delivery.

Key words: chitosan; sodium diclofenac; microparticles; ionic interaction

INTRODUCTION

Sodium diclofenac is a non-steroidal antiinflammatory drug (NSAID) used to relieve the inflammation, swelling, stiffness, and joint pain associated with rheumatoid arthritis, osteoarthritis, and ankylosing spondylitis. The main side effects of the drug (in case of the long-term use) may include: abdominal bleeding, pain or cramps, and swelling, anemia, blood clotting problems, constipation, diarrhea, dizziness, fluid retention, gas, headache, heartburn, indigestion, itching, nausea, peptic ulcers, rash, ringing in the ears, and vomiting. Sodium diclofenac remains un-dissolve in the acidic environment such as the stomach (pH 1 to 3) that leads to delay in the onset of drug effect, but

*Corresponding author's Email:

bsharifmakhmalzadeh@yahoo.com Telephone: +98-611-3373747 dissolves quickly in fewer alkaline conditions with pH 5-8 ^(1; 2). On the other hand, sodium diclofenac dissolves quickly in the intestinal alkaline condition so the effect of the long term drug uptake is impossible according to the short biological half life. Thus, total bioavailability of orally used sodium diclofenac has few important challenges.

So preparing formulations of diclofenac sodium with an immediate and controlled release property not only can increase competence and reduce the medication side effects ^(3,4). However, delayed in onset of the drug associated with very low biological half-life, causing repeated a day dose several times that has been both cause side effects and reduction in the patient compliance. In order to overcome these problems, a drug type can be design which improve the solubility in the stomach as well as reduce the onset time of the drug effect. These

will reduce dosage frequency following the slow controlled release. Therefore, the *efforts* of *many* researchers *have been* concerned to design a controlled release system for sodium diclofenac and other NSAID compounds ^(3,5, 6, 7).

Chitosan is a linear polysaccharide derived from chitin that has been used in different pharmaceutical forms and drug delivery systems ^(8, 9, 10, 11). Chitosan also were used in various membranes in order to simulate the human epidermis and animals such as mice and rabbits ^(12, 13). However, sodium diclofenac not only is a good candidate for the preparation of slow-release formulation, but the chitosan-sodium diclofenac prepared formula has several advantages include lower solubility in the intestine, slow-released form and improve drug absorption.

The purpose of this study was to prepare and evaluate the chitosan beads as a controlled drug release system for sodium diclofenac. Another purpose is to investigate the conditions in which the polymer bead is formed, and hence the dependence of the drug releases on bead formation.

Material and Methods Materials

Sodium diclofenac gift from was Hakim pharmaceutical Company (Tehran, Iran). Chitosan with the deacetylation degree of 95% and the molecular weight of 360 kDa was purchased from Fluka (Switzerland). Tween 80. sodium tripolyphosphate; monobasic potassium phosphate and sodium hydroxide were purchased from Merck (Germany). Dialysis bag was purchased from Toba Azema Co, Tehran (Iran). Minitab14 software was used for experimental design and the evaluation of the effect of variables on responses.

Determination of sodium diclofenac

The amount of microparticles loading and release rate of the drug determined using UV spectroscopy at wavelengths of 276 nm. The validity of assay method, linearity, repeatability, accuracy and limit of quantification (LOQ) were calculated.

Preparation of microparticles

Controlled release sodium diclofenac loaded microparticles were consisted of sustained and immediate released particles. In order to study the impact of independent variables on characteristics of microparticles, the full-factorial design with four variables and two levels were used. The percentage of Chitosan (%c), sodium poly phosphate (%s), Tween 80 (%T) and drug-chitosan proportion were selected as independent and drug loading efficiency, the rate of drug release and particle size as dependent variables (table 1). Sixteen formulations were prepared based on full factorial design. Sustained release microparticles prepared based on the method reported by Fattah et al ⁽¹⁴⁾.

Sodium diclofenac dispersed in water and pH was set on 6. Chitosan dissolved in 1% of acid acetic solution, pH adjusted on 5.2 then 1% of Tween 80 was added. Defined amount of drug solution was added dropwise to chitosan solution to providing desirable drug-chitosan proportion. Then a certain volume of chitosan-drug compound (5 ml) was added drop by drop with stirring into 10 ml of sodium polyphosphate solution. This mixture sonicated for 5 minutes and stored 24 hrs in refrigerator temperature. Microparticles were recovered by centrifugation at 4000 rpm for 5 min and dried at 50°C for 24 hrs.

The immediate particles were produced by poly vinyl prolidone (PVP) and manitol with equal proportion as carrier. PVP and manitol poured into container containing methanol until completely dispersed. Then the solvent was evaporated and stored in desiccators for two days and pulverized by

Behzad Sharif Makhmal Zadeh et al: Sodium Diclofenac Loaded Chitosan Controlled Release Microparticles

passing through 60-mesh sieve ⁽¹⁵⁾. Different controlled release formulations were obtained by

mixing sustained and immediate release particles with various ratios.

Independent veriable	Levels	
	Low (-)	High (+)
Percentage of chitosan	1%	3%
Percentage of sodium polyphosphate	6%	10%
Drug: lipid ratio	0.1	0.25
percentage of surfactant	0%	1%

Table 1: Independent variable and their levels in factorial experimental design

Drug entrapment efficiency

The percentage of encapsulated sodium diclofenac was determined by spectrophotometric determination at 276 nm after centrifugation of the aqueous dispersion. The amount of free drug was detected in the supernatant and the amount of encapsulated drug was calculated as a result of the initial drug minus the free drug. Amount of drug loading evaluated by two parameters; loading capacity (LC) and loading efficacy (LE) ⁽¹⁶⁾:

> LE= [(total amount of diclofenac) – (free diclofenac)]/ total amount of diclofenac LC= [(total amount of diclofenac) – (free diclofenac)]/ weight of microparticles

Morphological characterization

Surface morphology and stability of microparticles were studied by SEM (VP-1455, LEO, Germany). Microparticles were dried and coated by gold palladium.

Particle size analysis

The mean particle diameter and polydispersity index (PDI) of microparticles were calculated in water by laser light diffractometry, using Malvern MasterSizer SM 2000K (Malvern Instruments, UK). The samples were stirred and followed by sonication for 2 min for resuspending microparticles in water.

In vitro release study

Release studies for sixteen sustained formulation were performed in the simulated small intestine condition by pH 7.4. The release profile characterized using a six panel USP dissolution apparatus with 900 ml of medium with pH 7.4 and a temperature of 37°C. Furthermore, 5 ml of dissolution medium were withdrawn and replaced with fresh dissolution medium. Absorption of samples taken from centrifuge and separate clear supernatant solution and the appropriate dilution, by spectrophotometer at wavelength nm 276 against the control (dissolution test formulations were prepared without the drug). For immediate release particles, dissolution tests were performed same as sustained release microparticles in pH 2 and 7.4 for 2 hrs ⁽¹⁵⁾.

Results and Discussion

Validity of drug measurement method

The relationship between the light absorption and concentration was significant ($R^2 = 0.994$, P<0.001). Repeated surveys accountability in measurement methods within and between days for sodium diclofenac is represents the desired repeatability of measurement method on different days and caused nearly the same operation as well as error free

Int.J.Drug Dev. & Res., July-September 2010, 2(3): 468-475 Covered in Scopus & Embase results. The difference between real numbers and the estimated concentrations was about 5% that indicates the closeness of the estimated values to real results.

Encapsulation Efficiency

LE and LC of different sustained release microparticles are shown in table2. The correlation between LE and percentage of sodium polyphosphate was direct and significant (P=0.000), and percentage of Tween 80 (P= 0.000) was indirect and significant (P=0.006) correlation with drugchitosan proportion. This finding suggested physical mechanism for sodium diclofenac loading into chitosan microparticles. Chitosan is a polysaccharide with positive charge but sodium polyphosphate and sodium diclofenac in used pH demonstrate negative charges. In this state in ionic competition, increase in amount of sodium tripolyphosphate to cause reduction in drug loading but this was not observed. These results indicated physical mechanism as main mechanism for drug loading. This finding is contrary with results that suggested ionic interaction for sodium diclofenac loading in chitosan microparticles ⁽¹⁷⁾.

Piyakulawat et al ⁽⁶⁾ reported loading efficiency for sodium diclofenac - chitosan/carrageenan beads between 88-96% by that is higher than reported in present study.

On the other hand the correlation between independent variables and LC was not significant because the weight of microparticles were in wide range for all formulations. This finding indicated variation in microparticles collection.

Table 2: Loading efficiency (LE) and loading capacity (LC) of different sustained release microparticles (+ mean SD, n=3)

$(\pm \text{ mean SD}, \text{n}=3)$					
Batch number	Factorial design condition	LE	LC		
1	+ + - +	1.54 ± 80.34	1.22 ± 28.11		
2	+ + + -	1.39 ± 65.61	3.24 ± 26.19		
3	+ +	0.71 ± 66.05	1.14 ± 40.39		
4	+	1.5 ± 63.29	1.72 ± 41.27		
5	+ - + +	1.12 ± 57.39	0.99 ± 36.59		
6	+ +	0.59 ± 60.54	1.05 ± 37.10		
7	- + + -	1.33 ± 62.59	2.19 ± 30.19		
8	+ - + -	0/9 ± 50.74	1.80 ± 37.44		
9	- +	2.11 ± 71.49	2.71± 32.52		
10	+ -	0.71 ± 50.14	1.65 ± 36.52		
11		0.46 ± 54.12	2.37 ± 38.32		
12	- + + +	0.99 ± 77.05	1.66 ± 31.82		
13	+	0.67 ± 55.24	0.72 ± 39.23		
14	- + - +	1.29 ± 83.12	1.41 ± 35.39		
15	+ + + +	1.21±73.19	1.45 ± 28.94		
16	+ +	1.84 ± 70.54	2.19 ± 25.11		

Drug release

The result of drug dissolution in case of the sustained release particles is presented in table 3 as percentage at different time courses. Figure 1 illustrated the release profile. The findings show that only the correlation between tween 80 and drug

release after 1 and 8 hours was significant (P = 0.04 and P = 0.035 respectively). Percentage of drug released after 8 hours was in the range between 12 % from 95 %, which were observed in formulations 16 and 14, respectively. Based on the information obtained in this research increase in the tween 80 by

Behzad Sharif Makhmal Zadeh et al: Sodium Diclofenac Loaded Chitosan Controlled Release Microparticles

increasing the affinity of drug to chitosan reduced the released drug after 1 and 8 hours. Due to the nature of the chitosan and low hydrophilicity of sodium diclofenac, seems tween 80 increase the drugs loading and reduce the release percentage by reducing the surface tension and increase hydrophilicity.

On the other hand sodium polyphosphate percentage, although increased loading, but failed in affecting the percentage and speed of drug release. Sodium polyphosphate increases the LE through increasing the physical loading. Furthermore, because sodium diclofenac has no electrical interaction with chitosan, so the interaction between sodium phosphate and chitosan has no effect on the drug release rate. In site of the importance of electrical interactions between drug and chitosan, due to ion competition, with increasing concentration of sodium polyphosphate, release of drug from the polymer network should accelerate but such behavior was not observed. Drug release pattern that was observed from controlled release formulation was similar to sustained release particle, but higher amount of drug released after 1 and 8 hours was observed in case of controlled release formulation. Manjunatha et al reported 50 - 90 % of diclofenac released sodium from chitosan microparticles ⁽¹⁵⁾ which was in consistency with the result of drug release pattern in present study.

Table 3: Percentage of drug released after 1 (R_1) and 8 (R_8) hours from sustained release microparticles (mean \pm SD, n=3)

Batch number	Factorial design condition	R1	R8
1	+ + - +	23.83 ± 1.23	53.68 ± 3.71
2	+ + + -	14.08 ± .98	42.05± 3.25
3	+ +	22.07 ± 1.58	79.01 ± 4.17
4	+	7.9 ± .47	17.51 ± 1.96
5	+ - + +	15.76± 1.29	44.99 ± 2.63
6	+ +	28.21 ± 1.14	90.39 ± 6.11
7	- + + -	$26.92 \pm .72$	92.31 ± 8.19
8	+ - + -	20.72 ± 1.81	74.09 ± 4.88
9	- +	29.01± 2.60	84.38 ± 6.41
10	+ -	27.05 ± 2.03	93.17 ± 7.39
11		24.94 ± 1.50	74.19 ± 5.92
12	- + + +	23.91± 1.29	72.95± 7.01
13	+	31.80± 2.26	94.76 ± 8.51
14	- + - +	4.9 ± 0.44	11.73± 0.75
15	+ + + +	17.91± 1.33	62.14 ± 3.06
16	+ +	30.32 ± 2.89	95.4 ± 7.49

Int.J.Drug Dev. & Res., July-September 2010, 2(3): 468-475 Covered in Scopus & Embase



Figure 1: Rate of sodium diclofenac released from various sustained release formulations according to the time variable

Particle size distribution and morphology

The mean of particle size for sodium diclofenac loaded chitosan microparticles was found in the range of 4.8 (formulation 4) and 9.29 (formulation 12) μ m with monodisperse type. The relation between drug-chitosan proportion and particle size was direct and significant (P= 0.000). The morphology of chitosan microparticles as shown in Fig 2 indicates particles aggregation with irregular shape.



Fig 2: SEM picture of sodium diclofenac loaded chitosan microparticles

Int.J.Drug Dev. & Res., July-September 2010, 2(3): 468-475 Covered in Scopus & Embase

Conclusion

The sodium diclofenac loaded chitosan microparticles were prepared by ionic cross-linking with sodium tripolyphosphate successfully. Loading efficiency of sustained release microparticles to be touched by percentage of cross-linking agent, Tween 80 and drug-chitosan proportion. Chitosan microparticles provided suitable sustained release pattern in the manner that with increasing in percentage of Tween 80, the released amount reduced.

Acknowledgment

This paper is issued from Pharm D thesis and financial support was provided by Ahvaz Jundishapur University of Medical Sciences.

References

- Scholer, D.W., E.C., Ku, I., Boettcher, A., Schweizer, 1986 Pharmacology of diclofenac sodium. Am J Med. 80(4B):34-8. [PMID: 3085490]
- Schweitzer, A., N., Hasler-Nguyen, J., Zijlstra, 2009 Preferential uptake of the non steroid antiinflammatory drug diclofenac into inflamed tissues after a single oral dose in rats. BMC Pharmacol. 9:5. [PMID: 19291310]
- Gohel, M.C., and A.F. Amin, 1998 Formulation optimization of controlled release diclofenac sodium microspheres using factorial design. J Control Release. 51(2-3):115-22. [PMID: 9685908]
- 4) Barakat, N.S., A.A., Ahmad, 2008 Diclofenac sodium loaded-cellulose acetate butyrate: effect of processing variables on microparticles properties, drug release kinetics and ulcerogenic activity. J Microencapsul. 25(1):31-45. [PMID: 18188730]
- Gohel, M.C., and A.F., Amin, 1999 Formulation design and optimization of modified-release microspheres of diclofenac sodium. Drug Dev Ind Pharm. 25(2):247-51. [PMID: 10065360]
- 6) Piyakulawat, P., N., Praphairaksit, N., Chantarasiri, N., Muangsin, 2007 Preparation

and evaluation of chitosan/carrageenan beads for controlled release of sodium diclofenac. AAPS PharmSciTech. 8(4):E97. [PMID: 18181557]

- Avachat, A., and V., Kotwal, 2007 Design and evaluation of matrix-based controlled release tablets of diclofenac sodium and chondroitin sulphate. AAPS PharmSciTech. 8(4):E88. [PMID: 18181548]
- Zhang, H., I.A., Alsarra, S.H., Neau, 2002 An in vitro evaluation of a chitosan-containing multiparticulate system for macromolecule delivery to the colon. Int J Pharm. 239(1-2):197-205. [PMID: 12052705]
- 9) Georgeta, M., A.J., Elie, L., Didier, P., Luc, C., Adrian, M., Guy, 2004 Synthesis of chitosan microspheres containing pendant cyclodextrin moieties and their interaction with biological active molecules. Curr Drug Deliv. 1(3):227-33. [PMID: 16305386]
- 10) du Plessis, L.H., J., Lubbe, T., Strauss, A.F., Kotzé, 2010 Enhancement of nasal and intestinal calcitonin delivery by the novel Pheroid fatty acid based delivery system, and by N-trimethyl chitosan chloride. Int J Pharm. 385(1-2):181-6. [PMID: 19854253]
- 11) Shu, X.Z., and K.J., Zhu, 2000 A novel approach to prepare tripolyphosphate/chitosan complex beads for controlled release drug delivery. Int J Pharm. 201(1):51-8. [PMID: 10867264]
- 12) Rana, V., K., Babita, D., Goyal, A., Tiwary, 2004 Sodium citrate cross-linked chitosan films: optimization as substitute for human/rat/rabbit epidermal sheets. J Pharm Pharm Sci. 8(1):10-7. [PMID: 15946593]
- 13) Dureja, H., Tiwary, A.K., Gupta, S., 2001 Simulation of skin permeability in chitosan membranes. Int J Pharm. 213(1-2):193-8. [PMID: 11165107]
- 14) Fatah, EA., Grant, Gabr, KE., 1998. Physical characteristics and release behavior of salbutamol sulfate beads prepared with different ionic polysaccharides. Drug Develop Ind Pharm.24:541-7.
- 15) Manjunatha, K.M., Romana, M.V., Satyanarayana, D. 2007 Design and evaluation of

diclofenac sodium controlled drug delivery systems. Ind J Pharm Sci. 69: 384-389.

- 16) Lubben, I.M., Verhoef, J.C., Aelst, A.C., Borchard, G., Junginger, H.E. 2001 Chitosan microparticles for oral vaccination: preparation, characterization and preliminary in vivo uptake studies in murin payer patches. Biamat, 22: 687-694.
- 17) Boonsingrit, Y., Mitrerej, A., Muller, B.M. 2006 Chitosan drug binding by ionic intraction. Eur J. of Pharm and Biopharm,62:267-274.

Article History:-----Date of Submission: 20-02-10 Date of Acceptance: 12-05-10 Conflict of Interest: None Source of Support: Nil