

International Journal of Drug Development & Research | July-September 2011 | Vol. 3 | Issue 3 | ISSN 0975-9344 | Available online http://www.ijddr.in Covered in Official Product of Elsevier, The Netherlands ©2010 IJDDR

Evaluation of cardio protective Activity of Methanolic Extract Of Solanum Nigrum Linn. in Rats.

BHATIA NITISH^{*1}, MAITI PARTHA PRATIM¹, KUMAR ABHINIT¹, TULI ATUL¹, ARA TASNEEM², KHAN MASIH UZZAMAN¹

¹Sri Sai College of Pharmacy, Badhani, Pathankot, India; ²Drug Testing Laboratory, Dalgate, Srinagar, India.

Abstract

The present study was carried out to evaluate the cardioprotective and anti-oxidant activity of methanolic extract of berries of the family plant Solanum *nigrum* belonging to solanaceae. The cardioprotective activity of the extract was evaluated by using global in-vitro ischemia-reperfusion injury and tissue biochemical anti-oxidant profile respectively. The study was carried out using doses of 2.5 and 5.0 mg/kg for 6 days per week for 30 days. The results indicate that the extract exhibited significant (p < 0.001)cardioprotective activity against global in-vitro ischemia-reperfusion injury. The extract also exhibited significant (p<0.001) antioxidant potential as evident from the cardiac tissue biochemical antioxidant profile. Overall, the activities occurred in a dose-independent manner. The present study demonstrated that the methanolic extract of berries of the plant *Solanum* nigrum possessed cardioprotective and anti-oxidant activity and confirmed the traditional claims.

*Corresponding author, Mailing address: Mr. Nitish Bhatia Assistant Professor (Pharmacology) Sri Sai College of Pharmacy, Badhani Email: nitishnitish_18@yahoo.com Mobile: 09779709603

Key words:

Solanum nigrum, Cardioprotective, Anti-oxidant, fruit extract.

How to Cite this Paper:

Bhatia Nitish, Maiti Partha Pratim, Kumar Abhinit, Tuli Atul, Ara Tasneem, Khan Masih Uzzaman, "Evaluation Of Cardioprotective Activity Of Methanolic Extract Of *Solanum Nigrum* Linn. In Rats".Int. J. Drug Dev. & Res., Jul-Sep 2011, 3(3): 139-147

Copyright © **2010 IJDDR, Nitish Bhatia et al.** This is an open access paper distributed under the copyright agreement with Serials Publication, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Article History:-----Date of Submission: 24-04-2011 Date of Acceptance: 21-06-2011 Conflict of Interest: NIL Source of Support: NONE

Introduction

Coronary artery disease (CAD) is a leading cause of death in the Western world. Emergence of

ndia as a fast developing economy has also resulted in development of "office culture" characterized by Covered in Index Copernicus with IC Value 4.68 for 2010 **ULL Length Research Paper**

sedentary life style which is one of the key implications of CAD. The prevalence of CAD in urban North India varies from 7% to 10% [1] compared to 3% in USA [2] and <1% in Japan [3]. The CAD rates in South India are twofold higher than in North India, with Kerala reporting 14% in urban [4] and 7% in rural Thiruvananthapuram [5]. The treatment of IHD involves expensive and chronic drug therapy or equally expensive interventional procedures such as thrombolytic therapy and surgical recanalization [6], which has its own drawbacks in the form of reperfusion injury [7]. The etiopathogenesis of this phenomenon is complex and multifactorial of which oxygen free radicals (OFR) have been identified as the major contributor [8]. Efforts to contain OFR induced damage, using modern pharmacological agents have met with little success. Awareness of the rising incidence of ischemic heart disease (IHD) in India, coupled with prohibitive cost of treatment, particularly for developing countries has generated urgency for the rapid development of an alternative therapeutic modality which has always been available to us in the form of traditional or folklore medicine. Solanum nigrum Linn. (Solanaceae) is commonly known as 'Black nightshade'. The plant has been extensively used in traditional medicine in India and other parts of world to cure liver disorders [9], chronic skin ailments (psoriasis and ringworm) [10], inflammatory conditions, menstrual pain, fevers, diarrhea, eye diseases, hydrophobia etc. Keeping in view the need for a systematic scientific evaluation of medicinal plants in the amelioration of IHD, the present study was aimed at evaluating the cardioprotective potential of Solanum nigrum using global in-vitro ischemia-reperfusion injury as the animal model.

MATERIALS AND METHODS Plant materials

The plant Solanum nigrum Linn was collected from Haldia, Purba Medinipur Dt, during the month of March and April. That was identified by Dr. K. Gauthaman M.Pharm, Ph.D, Department of Pharmacognosy, Himalayan Pharmacy Institute, Majhitar, Sikkim, India. The berries were taken and dried in shade for 45 days. Then the shade-dried berries were made into coarse granules and were used for further investigation.

Experimental Animals

Male Albino rats (250-350 gm) were housed in the departmental animal house at an ambient temperature of 25°C, under a 12hour dark -12 hour light, cycle, for the whole period of the study (i.e. twelve weeks) and were fed with standard rat chow from Amrut Laboratory Animal feed, Bangalore and water, *ad libitum*. Experiments were carried out as per guidelines of Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), Ministry of Environment and Forest, Government of India(Reg. No.- 37/ 2007/ CPCSEA).

Chemicals & Reagents

All the chemicals and reagents used for these studies were purchased from Sigma chemicals, USA and SD fine chemicals, India.

Preparation of Extract

Dried fruit's coarse granules (850 gram) were divided in three parts, treated each three times with fresh methanol (1000 ml) separately for 48 h. The methanolic extracts thus obtained were combined, filtered and distilled on a water bath. The last traces of the solvent were evaporated under reduced pressure in rotatory evaporator (Heidolph Laborota 4011 digital). The yield of the methanolic extract was 6.75 % w/w (58 gram). Pharmacological studies were carried out by suspending a weighed amount of the extract in normal saline (95 ml): tween 80 (5 ml) ratio.

Determination of the test doses

A pilot level study was performed prior to initiation of protocol involving toxicological dose evaluation of alcoholic extract of *Solanum nigrum* in a dose range starting from 1 mg/Kg *p.o.* Mild toxic effects were observed at a dose of 7 mg/Kg *p.o.* Therefore, two median doses viz. 2.5 mg/Kg *p.o* and 5.0 mg/Kg *p.o* were selected for the present study.

Determination of cardioprotective activity

The rats were anaesthetized with ether, the chest opened and the heart along with one cm of ascending aorta attached was quickly removed and dipped in ice-cold saline. The hearts were then mounted on Langerdorff's apparatus (AD Instruments Australia) and perfused with Kreb's Hensleitt's (K-H) buffer [11] at a constant pressure of 60-70 mm Hg at 37°C, and aerated with a mixture of O_2 (95%) and CO_2 (5%). Following an initial period of 5 min of stabilization, the flow was stopped for 9 minutes (ischemia) followed by perfusion with K-H buffer for 12 minutes (Reperfusion) [12, 13]. At the end of ischemiareperfusion, the hearts were cut into two parts and one part was kept at -20°C for TBARS estimation and the other part was kept at -80°C in liquid nitrogen, till they were taken up for the estimation of SOD, reduced glutathione and tissue catalase content.

Estimation of Biochemical Parameters

Hearts from ten rats of each group were harvested and stored in liquid nitrogen for estimations of basal endogenous antioxidants. Roughly 0.5-1.0 gm of tissue was quickly cut out from the previously marked out area after reperfusion and immediately frozen in liquid nitrogen. The whole process was done in 8-10 seconds. The samples for biochemical estimations were weighed in an electronic pan balance and subsequently processed for the estimation of following parameters:

Myocardial Thiobarbituric Acid Reactive Substances (TBARS)

TBARS levels in the myocardium were determined by a modified version of the method described by

Ohkawa *et al* [14]. Results were expressed as nmol/g wet tissue wt.

Myocardial Reduced Glutathione [GSH]

Glutathione was estimated by the method described by Ellman *et al* [15]. Results were expressed as μ mole/g wet tissue weight.

Estimation of SOD

SOD levels in the hearts were determined by the method of McCord and Firdovich [16] and modified by Kakkar *et al* [17]. Results were expressed as IU/mg protein.

Myocardial Catalase

Catalase estimation for the tissue sample was done by the method of Aebi, [18]. Results were expressed as IU/mg protein

Histopathlogical Study

The myocardial tissue of rat from each group was taken and stained with Haematoxylin-Eosin (H-E) stain. After washing, the tissue was mounted on the compound microscope and myofibrillar structures were observed.

Statistical Analysis

Values are expressed as mean \pm SEM; significance is set at P \leq 0.05. One Way Analysis of variance (ANOVA) was carried out to test the significance of the biochemical data of the different groups in myocardial tissue samples.

Results

Table 1: Changes in myocardial tissue Thiobarbituric acid reactive substances Reduced (TBARS), Glutathione (GSH), Superoxide Desmutase (SOD) and Catalase (CAT) of the normal (N), Ischaemia reperfusion (I/R), and Ischaemia Reperfusion followed by Solanum nigrum extract (D1 = 2.5 mg/Kg p.o; D2 = 5.0 mg/Kg p.o), treated and Ischaemia Reperfusion followed by vehicle (I/R + V)treated groups. Results are represented as the mean \pm S.E.M. with n = 10 in each group. ^a p<0.05, as compared to the N group; $^{\rm b} p < 0.05$, as compared to I/R group; ^c *p*<0.05, as compared to D1 group

Covered in Index Copernicus with IC

Value 4.68 for 2010

FULL Length Research

Paper

Nitish Bhatia et al: Evaluation Of Cardioprotective Activity Of Methanolic Extract Of Solanum Nigrum Linn. In Rats

	Parameters			
Groups	TBARS	GSH	SOD	CATALASE
	nmole/g wet wt	μg/g wet wt	I.U/mg protein	I.U/mg protein
Ν	46.6 ± 1.2	378.5 ± 2.3	2.9 ± 0.1	43.0 ± 2.6
I/R	108.8 ± 5.5^{a}	294.3 ± 5.3^{a}	1.6 ± 0.2^{a}	27.4 ± 2.4^{a}
I/R + D1	88.4 ± 1.8^{b}	337.7 ± 5.8^{b}	$2.1 \pm 0.2^{\mathrm{b}}$	36.4 ± 1.3^{b}
I/R + D2	74.8 ± 4.7 ^{b, c}	357.4 ± 2.3^{b}	2.6 ± 0.3^{b}	$43.2 \pm 2.1^{b, c}$
D1 per se	52.5 ± 1.7	387.4 ± 3.2	2.8 ± 0.3	47.4 ± 1.2
D2 per se	69.2 ± 3.4^{a}	443.4 ± 5.3^{a}	2.9 ± 0.2	54.5 ± 1.3^{a}
I/R + V	107.9 ± 3.0^{a}	296.3 ± 2.8^{a}	1.4 ± 0.3^{a}	28.2 ± 1.4^{a}

Muocardial TBARS

Ischemia-Reperfusion injury resulted in significant increase in the levels of myocardial TBARS (108.8 \pm 5.5 nmoles/g wet wt; p < 0.001) as compared to the normal group (46.6 \pm 1.2 nmoles/g wet tissue wt). Significant increase in the TBARS levels was also seen in the I/R + vehicle group as well as D_2 per se group (107.9 ± 3.0 and 69.2 ± 3.4 nmoles/g wet tissue wt respectively). Treatment with methanolic extract of solanum nigrum (2.5 mg/Kg and 5.0 mg/Kg) significantly attenuated Ischemia-Reperfusion injury induced increase in myocardial tissue TBARS levels (88.4 \pm 2.8 and 74.8 \pm 4.7 nmoles/g wet tissue wt respectively)

Myocardial GSH

Ischemia-Reperfusion injury resulted in significant decrease in the levels of myocardial GSH (294.3 ± 5.3 μ g/g wet tissue wt; p < 0.001 respectively) as compared to normal group $(378.5 \pm 2.3 \ \mu g/g \text{ wet})$ wt.). Significant increase in the myocardial GSH levels was also seen in the $D_2 per se$ group (443.4 ± $6.4 \mu g/g$ wet tissue wt respectively) as compared to normal group (378.5 \pm 2.3 μ g/g wet wt.). Treatment with methanolic extract of solanum nigrum (2.5 mg/Kg and 5.0 mg/Kg) significantly attenuated Ischemia-Reperfusion injury induced decrease in myocardial tissue GSH levels $(337.7 \pm 5.8 \text{ and } 357.5)$ \pm 2.4 µg/g wet tissue wt respectively).

Myocardial Superoxide dismutase (SOD)

Ischemia-Reperfusion injury resulted in significant decrease in the levels of myocardial superoxide dismutase (1.6 \pm 0.2 I.U/mg protein) as compared to normal group $(2.9 \pm 0.1 \text{ I.U/mg proteins})$. Treatment with methanolic extract of solanum nigrum (2.5 mg/Kg and 5.0 mg/Kg) significantly attenuated Ischemia-Reperfusion injury induced decrease in myocardial tissue SOD levels (2.1 ± 0.2) and 2.6 ± 0.3 I.U/mg protein respectively)

Myocardial Catalase

Ischemia-Reperfusion injury resulted in significant decrease in the levels of myocardial catalase (27.4 \pm 2.4 I.U/mg protein) as compared to normal group (43.0 ± 2.6 I.U/mg protein). Treatment with methanolic extract of solanum nigrum (2.5 mg/Kg and 5.0 mg/Kg) significantly attenuated Ischemia-Reperfusion injury induced decrease in myocardial tissue catalase levels (36.4 \pm 1.3 and 43.2 \pm 2.1 I.U/mg protein respectively). Significant increase in the myocardial catalase levels was also seen in the D_2 per se group (54.5 ± 2.1 I.U/mg protein) as compared to normal group (43.0 ± 2.6 I.U/mg protein).



Slide 1: Normal rat myocardium showing well maintained myofibrillar structures.

Slide 2: Myocardium of the Rat subjected to I-R injury showing extensive degeneration of myofibrils with leukocytic accumulation, Edema and vacuolization.



3: Myocardium of the rat treated with low dose (2.5 mg/Kg) methanolic extract of *Solanum nigrum per se*.



Slide 4: Myocardium of the rat treated with high dose (5.0 mg/Kg) methanolic extract of *Solanum nigrum per se.*



Slide 5: Myocardium of the rat subjected to I-R injury and treated with low dose (2.5 mg/Kg) of methanolic extract of *Solanum nigrum*.



Slide 6: Myocardium of the rat subjected to I-R injury and treated with high dose (5.0 mg/Kg) of methanolic extract of *Solanum nigrum*.

Discussion

In the present study, Ischemia-Reperfusion injury resulted in an increase in the oxidative stress as evidenced by increase in myocardial TBARS and depletion of myocardial endogenous antioxidants enzymes (GSH. SOD and CAT). Chronic administration of the methanolic extract of Solanum nigrum at both the employed doses protected the hearts from ischemia-reperfusion induced oxidative stress, as evidenced by conservation of endogenous antioxidant enzyme levels and prevention in rise of TBARS to a significant extent. Ischemic-reperfusion injury is commonly associated with various stages of ischemic heart disease, starting from stable angina to acute myocardial infarction with spontaneous or

induced reperfusion [19] with oxidative stress playing a central role in its etiopathology of this condition [20]. This suggests that chronic administration of the extract imparted better oxidant stress bearing capacity in the rat hearts, by enhancing endogenous antioxidant.

Chronic *per se* oral administration of alcoholic extract of *Solanum nigrum* (at a dose of 5.0 mg/Kg) caused an increase in basal GSH and

CAT levels which indicated its possible adaptogenic property. However, one unique finding of the present study is that per se administration of alcoholic extract of Solanum nigrum was associated with a concominant rise in the TBARS levels. Although rise in TBARS is indicative of increased oxidative stress, it did not cause any overt cellular injury, as evidenced by histopathological study. It is, therefore possible that the increase in TBARS level was non-lethal and might be responsible for cellular adaptive mechanisms, leading to increased synthesis of endogenous antioxidants. Because, previously it has been reported that low levels of oxidative stress, cytokines, endo-toxins stimulate the synthesis of cellular antioxidants [21, 22, 23]. The major chemical compounds, identified in the alcoholic extract of Solanum nigrum were flavonids and glycosides and have significant antioxidants properties [24, 25]. However, many investigators are of the opinion that any antioxidant compound can also behave as a pro-oxidant [26, 27]. In this regard, melatonin has been shown to exert both pro- and antioxidant effects depending on the concentration and duration of exposure [28]. Augmentation of basal endogenous antioxidant reserve by any therapeutic agent is presently the focus of great scientific interest [29, 30] as it is expected to offer better protection against oxidative stress than any exogenously administered antioxidants. It has also been shown that transgenic mice overexpressing CuZn-SOD are more resistant to cardiac injury, than mice, which are exogenously administered with SOD [31]. Moreover, increase in GSH and catalase, as observed in the present study, is supposed to be more beneficial than SOD, because increase in SOD without a concomitant increase in catalase may lead to intracellular accumulation of H_2O_2 [32]. Among medicinal plants, similar effects have been observed with chronic administration of garlic [33]. In the present study the isolated perfused rat heart as a model was used to investigate the cardioprotective action of *Solanum nigrum*, and the choice of the preparation is very wide and best characterized. This model was also the most frequently used one for the development and assessment of anti ischemic agents.

Chronic oral administration of alcoholic extract of Solanum nigrum in rat augments myocardial endogenous antioxidants, without causing any cellular injury. It has a definite, dose-dependent modulatory effect on the antioxidant milieu of heart. The drug treated groups offered better cardioprotection when subjected to IR injury. There was also no definite pattern of change in the level of TBARS and the endogenous antioxidants. Hence these modulatory actions seem to be enzymestudy specific. The reveals an important cardioprotective action of the Solanum nigrum, a widely used Indian medicinal plant in IHD.

The present study reveals important cardioprotective of action of the alcoholic extract of *Solanum nigrum,* a widely used Indian medicinal plant in ischemic heart disease.

Acknowledgement

The authors would like to heartily thank Er. S.K Punj, Chancellor, Sri Sai University, Palampur and Mrs. Tripta Punj, M.D, Sri Sai Group of Institutions, Badhani, Pathankot for their unending support and inspiration towards this research work.

References

 Reddy KS. Cardiovascular diseases in India. World health Stat Q. 1993; 46: 101-107.

- DeStefano F, Merritt RK, Anda RF, Casper ML, Eaker ED. Trends in nonfatal coronary heart disease in the United States, 1980 through 1989. Arch. Intern. Med 1993; 153(21): 2489-2494.
- Verschuren WM, Jacobs DR, Bloemberg BP, Kromhout D, Menotti A, Aravanis C, Blackburn H, Buzina R, Dontas AS, Fidanza F. Serum total cholesterol and long-term coronary heart disease mortality in different cultures. Twentyfive-year follow-up of the seven countries study. JAMA 1995; 274(2): 131-136.
- 4. Singh RB, Niaz AM, Ghosh S, Agarwal P, Ahmad S, Begum R, Onouchi Z, Kummerow FA. Randomized, controlled trial of antioxidant vitamins and cardioprotective diet on hyperlipidemia, oxidative stress, and development of experimental atherosclerosis: the diet and antioxidant trial on atherosclerosis (DATA). Cardiovasc. Drugs Ther 1995; 9(6): 763-771.
- Kutty VR, Balakrishnan KG, Jayasree AK, Thomas J. Prevalence of coronary heart disease in the rural population of Thiruvananthapuram district, Kerala, India. Int J Cardiol 1993; 39(1): 59-70.
- Herrin J, Cangialose CB, Boccuzzi SJ, Weintraub WS, Ballard DJ. Household income losses associated with ischaemic heart disease for US employees. Pharmacoeconomics 2000; 17(3): 305-314.
- 7. Ohnishi Y, Butterfield M, Saffitz JE, Sobel BE, Corr PB, Goldstein JA.
 Deleterious effects of a systemic lytic state on reperfused myocardium.
 Minimization of reperfusion injury and enhanced recovery of myocardial

function by direct angioplasty. Circulation 1995; 92(3): 500-510.

- Elahi MM, Kong YX, Matata BM. Oxidative stress as a mediator of cardiovascular disease. Oxid Med Cell Longev 2009; 2(5): 259-269.
- Hsieh CC, Fang HL, Lina WC. Inhibitory effect of Solanum nigrum on thioacetamide-induced liver fibrosis in mice. J Ethnopharmacol 2008; 119: 117-121.
- Ahmed AH, Kamal IH, Ramzy RM. Studies on the molluscicidal and larvicidal properties of Solanum nigrum L. leaves ethanol extract. J Egypt Soc Parasitol 2001; 31: 843-852.
- Borchgrevink PC, Jynge P. Direct effects of furosemide and amiloride on the perfused and ischaemic rat heart. Pharmacol Toxicol 1989; 64(1): 100-106.
- Maulik SK, Kumari R, Maulik M, Reddy KS, Seth SD. Effect of flavone in a canine model of myocardial stunning. Indian J Exp Biol 1999; 37(10): 965-970.
- 13. Gauthaman K, Maulik M, Kumari R, Manchanda SC, Dinda AK, Maulik SK. Effect of chronic treatment with bark of Terminalia arjuna: a study on the isolated ischemic-reperfused rat heart. J Ethnopharmacol 2001; 75(2-3):197-201.
- 14. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal Biochem 1979; 95(2): 351-358.
- Ellman GL. Tissue sulfhydryl groups. Arch Biochem Biophys 1959; 82(1): 70-77.
- McCord JM, Fridovich I. The utility of superoxide dismutase in studying free radical reactions. I. Radicals generated

Nitish Bhatia *et al:* Evaluation Of Cardioprotective Activity Of Methanolic Extract Of *Solanum Nigrum* Linn. In Rats

y the interaction of sulfite, dimethyl sulfoxide, and oxygen. J Biol Chem 1969; 244(22):6056-63.

υ

- 17. Kakkar P, Das B, Viswanathan PN. A modified spectrophotometric assay of superoxide dismutase. Indian J Biochem Biophys 1984; 21(2): 130-132.
- Aebi H. Method in Enzymatic Analysis Vol 3. NewYork, USA, Academic Press Inc, 1974, pp. 673–686.
- Gross GJ, Kersten JR, Warltier DC. Mechanisms of postischemic contractile dysfunction. Ann Thorac Surg 1999; 68(5): 1898-1904.
- 20. Bolli R. Why myocardial stunning is clinically important. Basic Res Cardiol 1998; 93(3):169-72.
- 21. Bensard DD, Brown JM, Anderson BO, Banerjee A, Shanley PF, Grosso MA, Whitman GJ, Harken AH. Induction of endogenous tissue antioxidant enzyme activity attenuates myocardial reperfusion injury. J Surg Res 1990; 49(2): 126-131.
- 22. Singal PK, Siveski-Iliskovic N, Hill M, Thomas TP, Li T. Combination therapy with probucol prevents adriamycininduced cardiomyopathy. J Mol Cell Cardiol 1995; 27(4): 1055-1063.
- 23. Maulik N, Yoshida T, Das DK. Oxidative stress developed during the reperfusion of ischemic myocardium induces apoptosis. Free Radic Biol Med 1998; 24(5): 869-875.
- 24. Huk I, Brovkovych V, Nanobash Vili J, Weigel G, Neumayer C, Partyka L, Patton S, Malinski T. Bioflavonoid quercetin scavenges superoxide and increases nitric oxide concentration in ischaemia-

reperfusion injury: an experimental study. Br J Surg 1998; 85(8): 1080-1085.

- 25. Sugihara N, Arakawa T, Ohnishi M, Furuno K. Anti- and pro-oxidative effects of flavonoids on metal-induced lipid hydroperoxide-dependent lipid peroxidation in cultured hepatocytes loaded with alpha-linolenic acid. Free Radic Biol Med 1999; 27(11-12): 1313-1323.
- Diplock AT. Defense against reactive oxygen species. Free Radic Res 1998; 29(6): 463-467.
- 27. Halliwell B. Lipid peroxidation, antioxidants and cardiovascular disease: how should we move forward? Cardiovasc Res 2000; 47(3): 410-418.
- 28. Osseni RA, Rat P, Bogdan A, Warnet JM, Touitou Y. Evidence of prooxidant and antioxidant action of melatonin on human liver cell line HepG2. Life Sci 2000; 68(4): 387-399.
- 29. Maulik G, Maulik N, Bhandari V, Kagan VE, Pakrashi S, Das DK. Evaluation of antioxidant effectiveness of a few herbal plants. Free Radic Res 1997; 27(2): 221-228.
- 30. Pathania V, Syal N, Pathak CM, Khanduja KL. Changes in rat alveolar macrophageal antioxidant defense and reactive oxygen species release by high dietary vitamin E. J Nutr Sci Vitaminol (Tokyo) 1998; 44(4): 491-502.
- 31. Wang S, Zhu H, Chen C. Reactive oxygen species contribute to the induction of superoxide dismutase during heat shock in cultured rat neonatal cardiomyocytes. Chin Med J (Engl) 2000; 113(7): 606-609.
- 32. Yim MB, Chock PB, Stadtman ER.

opper, zinc superoxide dismutase catalyzes hydroxyl radical production from hydrogen peroxide. Proc Natl Acad Sci 1990; 87(13): 5006-5010.

33. Banerjee SK, Maulik M, Manchanda SC, Dinda AK, Das TK, Maulik SK. Garlicinduced alteration in rat liver and kidney morphology and associated changes in endogenous antioxidant status. Food Chem Toxiol 2001; 39(8): 793-797.



J

