In-vivo Screening of Analgesic and Antiulcer Activity on Carum carvi Seeds

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Abstract

The present study was designed to investigate the antiulcer and analgesic potential of Carum carvi seeds. caraway seeds use is common in diet, so there is no need for special administration of the drug and by little increase in intake quantity, the use of analgesics is abolished. Antiulcer activity was also evaluated by aspirin induced ulcer models. Effect of concurrent administration of ethanolic and aqueous extracts of seeds extract Carum carvi at a dose of 100 and 200 mg/kg b.w. respectively was given by oral route. Ethanolic and aqueous extracts of Carum carvi seeds significantly reduction in gastric content, total acidity, ulcer index, and increase in pH of gastric pylorus ligation ulcer model. In comparison with the standard drug, the results of hydro alcoholic extract at 100 mg dose showed good analgesic and at 200 mg dose showed antiulcer activity compared with a standard drug. Extracts of Carum carvi may be useful as a natural analgesic in the treatment of ulcer, inflammation, and pain.

Keywords: Carum carvi; Antulcer activity; Aspirin; Analgesic

Introduction

An analgesic or painkiller is any member of the group of drugs used to achieve analgesia, relief from pain. A number of herbal drugs are used to treat Analgesia. Several epidemiological studies from different countries have reported widely varying prevalence rates for chronic pain, ranging from 12 to 80% of the population [1-6]. It becomes more common as people approach death. A study of 4,703 patients found that 26% had pain in the last two years of life, increasing to 46% in the last month [2]. The intensity of chronic pain was higher for girls, and girls' reports of chronic pain increased markedly between ages 12 and 14 [3]. Perception gives information on the pain's location, intensity, and something about its nature. The various conscious and unconscious responses to both sensation and perception, including the emotional response, add further definition to the overall concept of pain [4]. Pain perception also varies depending on the location of the pain. The kinds of stimuli that cause a pain response on the skin include pricking, cutting, crushing, burning, and freezing. These same stimuli would not generate much of a response in the intestine. Intestinal pain arises from stimuli such as swelling, inflammation, and distension [4]. Pain is the main reason for visiting the emergency department in more than 50% of cases and is present in 30% of family practice visits [5]. Several epidemiological studies from different countries have reported widely varying prevalence rates for chronic pain, ranging from 12 to 80% of the population [6]. It becomes more common as people approach death. A study of 4,703 patients found that 26% had pain in the last two years of life, increasing to 46% in the last month [7]. The intensity of chronic pain was higher for girls, and girls' reports of chronic pain increased markedly between ages 12 and 14 [8]. The success of NSAIDs in treating various inflammatory conditions such as rheumatoid arthritis (RA) and osteoarthritis (OA) validated inhibition of the enzyme prostaglandin H synthase (PGHS) or cyclooxygenase (COX) as a highly suitable target in anti-inflammatory therapies [9,10]. The following NSAIDs are derived from fenamic acid. Which is a derivative of anthranilic acid [11], which in turn is a nitrogen isostere of salicylic acid, which is the active metabolite aspirin [11]. The first purified preparation of COX enzyme was reported in 1976 [12]. More than a decade later COX was cloned in 1988 [13-15]. In the 1990’s an inducible isoform now called COX-2 was discovered [16-18]. Inflammation (Latin, inflammation) is part of the complex biological response of body tissues to harmful stimuli, such as pathogens, damaged cells, or irritants [19-32].

Caraway (Carum carvi), also known as meridian fennel, [19-22] or Persian cumin, [31,33] is a biennial plant in the family Apiaceae, [23] native to western Asia, Europe, and northern Africa. It occurs in India to a limited extent in the temperate regions of the western Himalayas, Jammu and Kashmir and in chakratha hills of utter Pradesh. however, polyploid variants (with four haploid sets=4 n) of this plant that was found to be perennial. Finland supplies approximately 28% (2011) of the world’s caraway production [24]. Caraway seeds are a rich source of dietary fiber. 100 g seeds provide 38 g of fiber,100% of daily recommended intake of fiber. The fruits, usually used whole, have a pungent, anise-like flavor and aroma that comes from essential oils, mostly carvone, limonene and anethole [25,26]. It is used in caraway seed cake, and it is frequently added to sauerkraut [27-30]. The roots may be cooked as a vegetable like parsnips or carrots. Additionally, the leaves are sometimes consumed as herbs, either raw, dried, or cooked, similar to parsley [31-38].

Materials and Methods

Extraction and preparation of test sample

The seeds of caraway were collected, dried and ground into fine powder. 250 grams of caraway powder was weighed and extracted by using two types of solvents. Extraction was carried out by using 500 ml of aqueous and hydroalcohol respectively by using maceration process. Until the complete extraction takes place. The extract obtained was filtered by using vacuum filtration and further, concentrated by using evaporation. The extract was further dried under air.

Albino Wistar strain rats obtained from Mahaveera enterprises, peerzadiguda, ghatkesar mandal, R.R. Dist-39. weighing 150-250 gm of either sex was used in this study. Animals were acclimatized to laboratory conditions before testing e.g., room temperature of 25 ±
The experimental protocol was approved by the Institutional Animal Ethics Committee (IAEC) and protocol approval No. is 011/IAEC/Ncpa/Researchproposal/2015/16. The committee for purpose of control and supervision of experiments on animals (CPCSEA) Reg. No. is 1629/poa/12/CPCSEA. All the experimental procedures were carried out according to the proper guidelines of CPCSEA.

**Analgesic studies**

Tail immersion method: In present study analgesia was assessed according to the method of Luiz et al. The animals fasted for 12 hours before the procedure was started. A total of six rats was grouped into four groups. One group treated with distilled water serves as control. The second group was treated Diclofenac sodium which acts as an standard. Other two groups were given test drugs i.e, caraway extract of 100 mg/kg and 200 mg/kg. The drug is administered through oral route by using oral feeding needle. They were held in position in a suitable restrainer with a tail extending out. 3-4 cms of the tail were marked and immersed in water bath thermostatically maintained at 55°C. The withdrawal time of tail from hot water (in seconds) was noted as reaction time or tail flick latency. The maximum cutoff time for immersion was 15 seconds to avoid the injuries of tissues of the tail. The initial reading was taken immediately before administration of the test and standard drugs and then 15,30,45,60 and 75 minutes after the administration.

Anti-ulcer screening method: Anti-ulcer activity was evaluated by an aspirin-induced ulcer in Wistar rats [8].

Aspirin-induced gastric ulcer model: The animals fasted for 24 hours before the experiment. Animals were divided into four groups each containing 6 animals. Group 1 served as a positive control and received omeprazole at the dose of 30 mg/kg body weight, group 2 served as a negative control received aspirin at the dose of 500 mg/kg body weight. Group 3 received the aqueous extract of caraway seeds at the dose of 500 mg/kg body weight and group 4 received the hydroalcoholic extract of caraway seeds at the dose of 500 mg/kg body weight. After one hour of extract administration, aspirin at a dose of 500 mg/kg body weight was administered orally to all four groups. Then after 6 hours animals were euthanized with an excess of chloroform and theRomach, animals were killed, stomach was removed and the contents were drained into graduated centrifuge tube. The stomach was removed and the contents were drained into graduated centrifuge tube through a small nick along the greater curvature. The volume of the juice was measured.

Induction of gastric ulcer: The animals were fasted for 36 hours in separate cages with raised wide wire mesh to avoid coprophagia (Basso et al.), but with water given ad libitum. After 4 hours under light anesthesia by ketamine, animals were sacrificed. The abdomen was opened and a ligature was placed around the esophagus. The stomach was removed and the contents were drained into graduated centrifuge tube. The stomach was opened on greater curvature and examined for ulceration. Evaluation of the degree of ulceration was expressed in terms of ulcer score which is calculated by dividing the total number of ulcers in each group by a number of rats in that group (Robert et al.). The degree of ulceration was also expressed as ulcer index and calculated by multiplying ulcer score × 100 (Radwan et al.).

**Acid volume**

Procedure: Under ether anesthesia, a midline incision is made. Herbal extract was given to rats as per dosages and after a span of 1 hour aspirin was given at a dose of 500 mg/kg body weight. 3 hours after ingestion of aspirin, the animals were sacrificed. The abdomen was opened and a ligature was placed around the esophagus. The stomach was removed and the contents were drained into graduated centrifuge tube. The volume of the juice was measured.

**Determination of total acidity and free acidity**

Procedure: 0.1 mL of gastric juice specimen was transferred in a porcelain evaporating dish. 1-2 drops of Töpler’s reagent is added. A colour change was observed; a bright red colour appears if free hydrochloric acid is present. 1-2 drops of phenolphthalein were added to the gastric juice with Töpler’s reagent. Titrated with 0.01 N NaOH from a burette, mixing was done after each addition until the last trace of red colour disappeared and was replaced by a canary yellow colour. The numbers of milliliters of NaOH used was read from the burette. This represents the amount of free hydrochloric acid. The titration was continued until the red colour of phenolphthalein appeared (deep pink), titrated to the point at which the further addition of alkali did not deepen the colour. Reading was taken (mL NaOH) for total acidity.

Calculation: \[ Y = mL \text{ of } 0.1N \text{ NaOH} \times 10 \]

where, \( Y = \text{Total acidity (mEq/L); Acidity} = \text{Volume of NaOH} \times \text{Normality} \times 100 \text{ mEq/litre} \)

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**Induction of gastric ulcer**: The animals were fasted for 36 hours in separate cages with raised wide wire mesh to avoid coprophagia (Basso et al.), but with water given ad libitum. After 4 hours under light anesthesia by ketamine, animals were killed, stomach was removed and opened on greater curvature and examined for ulceration. Evaluation of the degree of ulceration was expressed in terms of ulcer score which is calculated by dividing the total number of ulcers in each group by a number of rats in that group (Robert et al.). The degree of ulceration was also expressed as ulcer index and calculated by multiplying ulcer score × 100 (Radwan et al.).

Preventive index was calculated according to the method:

\[ \text{Preventive index} = \frac{\text{U-I control-U-I. treated}}{\times \times 100} \]

% of ulceration= Number of ulcerated rats-Number of non-ulcerated rats × 100/Number of rat in group

### Table 1: Results of aqueous extract of Carum carvi seeds by using tail immersion method.

<table>
<thead>
<tr>
<th>Time in minutes</th>
<th>At 0 min</th>
<th>At 15 min</th>
<th>At 30 min</th>
<th>At 45 min</th>
<th>At 60 min</th>
<th>At 75 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>2.5 ± 0.02</td>
<td>2.5 ± 0.02</td>
<td>2.55 ± 0.05</td>
<td>2.6 ± 0.05</td>
<td>2.5 ± 0.02</td>
<td>2.5 ± 0.02</td>
</tr>
<tr>
<td>Standard</td>
<td>2.5 ± 0.05***</td>
<td>6.2 ± 0.15***</td>
<td>8.4 ± 0.2***</td>
<td>12 ± 0.4***</td>
<td>9 ± 0.5***</td>
<td>5 ± 0.5***</td>
</tr>
<tr>
<td>Aqueous 100mg</td>
<td>2.6 ± 0.11ns</td>
<td>5.6 ± 0.04**</td>
<td>8.1 ± 0.07**</td>
<td>10.5 ± 0.1**</td>
<td>6.5 ± 0.09**</td>
<td>5.5 ± 0.11**</td>
</tr>
<tr>
<td>Aqueous 200mg</td>
<td>2.5 ± 0.1ns</td>
<td>6.3 ± 0.09ns</td>
<td>8.4 ± 0.1ns</td>
<td>12.5 ± 0.1**</td>
<td>10.3 ± 0.16**</td>
<td>8 ± 0.88**</td>
</tr>
</tbody>
</table>

Values are mean ± SD, n=6 rats in each group. Significant values calculated by dunnet’s multiple comparision test. Here p values, ***p: <0.001-Highly significant, **P: <0.01-Significant, p value: >0.05-Non-significant.
Results

Results of this study are presented in Tables 1–4 and Figures 1–5.

Phytochemical screening results

Phytochemical screening results are presented in Table 5.

Discussion

Analgesic studies

The time taken in seconds for the rat to withstand its tail in hot water at 55°C was noted. In control animals, without any drug, almost all rats withstand for approximately 2.5 sec at varying time intervals. The time taken for the standard group i.e., group of rats treated with standard drug Diclofenac sodium the time in seconds for the rat to withdraw its tail from hot water is increased with increase in time. At a maximum of 45 min after ingestion of standard drug the rat could withstand for 12 secs showing the analgesic effect. In comparison with standard drug Diclofenac sodium the time in seconds for the rat to withdraw its tail from hot water at 55°C was noted. In control animals, almost all animals are positive for an ulcer. Alkaloids, glycosides, carbohydrates, non-reducing sugars, steroids, aminoacids, saponins, volatile oils are absent in the screening of phytochemicals.

Phytochemical screening

Proteins, tannins, phenolic compounds and flavonoids are present.

Conclusion

Review literature supported the use of seeds of Carum carvi for Analgesic and Anti-Ulcer Activity. Extraction using aqueous and hydroalcoholic solvents was done. Screening of Analgesic activity is

Table 2: Results of hydro alcoholic extract of Carum carvi seeds by using tail immersion method.

<table>
<thead>
<tr>
<th>Time in min</th>
<th>At 0 min</th>
<th>At 15 min</th>
<th>At 30 min</th>
<th>At 45 min</th>
<th>At 60 min</th>
<th>At 75 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>2.5 ± 0.2</td>
<td>2.5 ± 0.2</td>
<td>2.5 ± 0.5</td>
<td>2.6 ± 0.05</td>
<td>2.5 ± 0.02</td>
<td>2.5 ± 0.02</td>
</tr>
<tr>
<td>Standard</td>
<td>2.5 ± 0.05***</td>
<td>6.2 ± 0.15***</td>
<td>8.4 ± 0.2***</td>
<td>12 ± 0.4***</td>
<td>9 ± 0.5***</td>
<td>5 ± 0.5***</td>
</tr>
<tr>
<td>Hydro alcoholic 100 mg</td>
<td>2.5 ± 0.07ns</td>
<td>4.4 ± 0.23**</td>
<td>6.4 ± 0.15**</td>
<td>10.2 ± 0.14**</td>
<td>6.2 ± 0.20**</td>
<td>5.1 ± 0.14ns</td>
</tr>
<tr>
<td>Hydro alcoholic 200 mg</td>
<td>2.5 ± 0.08ns</td>
<td>5.4 ± 0.09**</td>
<td>7.4 ± 0.15**</td>
<td>11.2 ± 0.17**</td>
<td>9 ± 0.37ns</td>
<td>5 ± 0.1ns</td>
</tr>
</tbody>
</table>

Values are mean ± SD, n=6 rats in each group. Significant values calculated by dunnet’s multiple comparison test. Here p values, ***p: <0.001-Highly significant, **P: <0.01-Significant, p value: >0.05-Nonsignificant

Table 3: Results of anti-ulcer activity using hydro-alcoholic extract of Carum carvi seeds against aspirin-induced gastric ulcers.

<table>
<thead>
<tr>
<th>Group</th>
<th>Volume of Centrifuge (ml)</th>
<th>Volume of NaOH Consumed (ml)</th>
<th>Total Acidity (mEq/L)</th>
<th>Ulcer Index</th>
<th>% Ulcer Protection</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1 ± 0.36</td>
<td>2.5ml ± 0.72</td>
<td>25 ± 2.56</td>
<td>6 ± 1.05</td>
<td>-</td>
</tr>
<tr>
<td>Standard</td>
<td>0.8 ± 0.34</td>
<td>1 ± 0.36</td>
<td>10 ± 0.45***</td>
<td>1.5 ± 0.51***</td>
<td>75 ± 3.57</td>
</tr>
<tr>
<td>Hydro alcoholic 100 mg</td>
<td>0.7 ± 0.37</td>
<td>2.1 ± 0.4</td>
<td>21 ± 1.96**</td>
<td>5 ± 0.7**</td>
<td>16.7 ± 0.37</td>
</tr>
<tr>
<td>Hydro alcoholic 200 mg</td>
<td>0.4 ± 0.58</td>
<td>1 ± 0.36</td>
<td>10 ± 0.47ns</td>
<td>2 ± 0.58**</td>
<td>66.7 ± 1.09</td>
</tr>
</tbody>
</table>

Values are mean ± SD, n=6 rats in each group. Significant values calculated by dunnet’s multiple comparison test. Here p values ***p: <0.001-Highly significant, **P: <0.01-Significant, p value: >0.05-Nonsignificant

Table 4: Results of anti-ulcer activity using aqueous extract of Carum carvi seeds against aspirin-induced gastric ulcers.

<table>
<thead>
<tr>
<th>Group</th>
<th>Volume of Centrifuge (ml)</th>
<th>Volume of NaOH Consumed (ml)</th>
<th>Total Acidity (mEq/L)</th>
<th>Ulcer Index</th>
<th>% Ulcer Protection</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1 ± 0.36</td>
<td>2.5ml ± 0.72</td>
<td>25 ± 2.56</td>
<td>6 ± 1.05</td>
<td>-</td>
</tr>
<tr>
<td>Standard</td>
<td>0.8 ± 0.34</td>
<td>1 ± 0.36</td>
<td>10 ± 0.45***</td>
<td>1.5 ± 0.51***</td>
<td>75 ± 3.57</td>
</tr>
<tr>
<td>Aqueous extract 100 mg</td>
<td>0.8 ± 0.36</td>
<td>1.8 ± 0.53</td>
<td>18 ± 0.58**</td>
<td>5.5 ± 0.52**</td>
<td>8.3 ± 0.38</td>
</tr>
<tr>
<td>Aqueous extract 200 mg</td>
<td>0.4 ± 0.58</td>
<td>1.3 ± 0.36</td>
<td>13 ± 0.55**</td>
<td>3 ± 0.48**</td>
<td>50 ± 0.51</td>
</tr>
</tbody>
</table>

Values are mean ± SD, n=6 rats in each group. Significant values calculated by dunnet’s multiple comparison test. Here p values ***p: <0.001-Highly significant, **P: <0.01-Significant, p value: >0.05-Nonsignificant
Table 5: Table showing results of phytochemical screening.

<table>
<thead>
<tr>
<th>S. No</th>
<th>Name of the Experiment</th>
<th>Hydro alcoholic extract</th>
<th>Hydro extract</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Detection of Alkaloids:</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>2</td>
<td>Test for Glycosides:</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>3</td>
<td>Test for cardiac glycosides:</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>4</td>
<td>Tests for deoxy sugars:</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>5</td>
<td>Tests for anthraquinone glycosides:</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>6</td>
<td>Tests for cyanogenetic glycosides</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>7</td>
<td>Test for saponin glycosides:</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>8</td>
<td>Test for coumarin glycosides:</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>9</td>
<td>Test for carbohydrates:</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>10</td>
<td>Test for non-reducing sugars:</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

Figure 1: Graph showing effect of aqueous extract on time in sec results by tail immersion method.

Figure 2: Graph showing effect of hydro alcoholic extract on time in sec results by tail immersion method.
done by using tail immersion method at a dose of 100 mg and 200 mg by using two extracts. In comparison with the standard drug, the results of hydroalcoholic extract at 100 mg dose showed good analgesic activity compared with a standard drug. Anti-Ulcer activity is screened by using Aspirin-induced ulcer at doses of 100 mg and 200 mg. At a dose of 200 mg of hydroalcoholic extract, the percentage of ulcer protection in test group is almost similar with ulcer protection with a standard drug. It decreases the total acidity and ulcer index. Finally, we conclude that hydroalcoholic extract at a dose of 100 and 200 mg shows good Analgesic and Anti-Ulcer Activity. The drug was selected for a future investigation involving column chromatographic separation of extracts and to isolate and identify the actual chemical constituent responsible for the activity.

References


Figure 5: Images showing ulcer index produced in animals during screening of anti-ulcer activity of two different solvent extracts of Carum carvi against aspirin-induced gastric ulcers in rats.