

## Research Article

# Antiepileptic Activity of Methanol Extract of *Butea monosperma* (Lam.) Kuntze and its Isoalted Bioactive Compound in Experimentally Induced Convulsion in Swiss Albino Mice

Manas Kumar Das<sup>1</sup>, Papiya Mitra Mazumder<sup>2</sup> and Sanjita Das<sup>3</sup>

<sup>1</sup>Department of Pharmacy, IEC College of Engineering and Technology, Greater Noida, Uttar Pradesh, India

<sup>2</sup>Department of Pharmaceutical Sciences and Technology, Birla Institute of Technology, Mesra, Ranchi, Jharkhand, India

<sup>3</sup>Department of Pharmaceutical Technology, Noida Institute of Engineering and Technology, Greater Noida, Uttar Pradesh, India

\*Corresponding author: Manas Kumar Das, Department of Pharmacy, IEC College of Engineering and Technology, Greater Noida, Uttar Pradesh, India, Tel: 01202326665; E-mail: manas.kd@gmail.com

Received November 26, 2015; Accepted January 10, 2016; Published January 15, 2016

## Abstract

The purpose of this research work is to establish the anticonvulsant activity of *Butea monosperma* (Lam.) Kuntze (Fabaceae) generally used in traditional medicine for the treatment of neurodegenerative diseases, amnesia and mental illness. In this study, the anticonvulsant activity of the crude methanol stem extract of *Butea monosperma* (BMME) and its isolated bioactive compound at doses of 100, 200 and 300 mg/kg and 20 mg/kg respectively using Pentylenetetrazole (PTZ) induced convulsion (chemically induced convulsion) and Maximal electroshock convulsion (MES) induced convulsion (electrically induced convulsion) models were investigated in Swiss albino mice. The sedative and hypnotic effect was also studied using the Phenobarbital induced sleep model. The onset of Phenobarbital induced sleep was decreased dose dependently, there was a marked increase in the duration of sleep when compared with the control group. BMME at doses of 100-300 mg/kg significantly decreased ( $p<0.001$ ) the tonic hind limb extension in MES induced convulsion. There was 66.67% and 83.33% protection against convulsion and mortality respectively at the highest dose of 300 mg/kg. In PTZ induced convulsion, BMME delayed all the parameters like onset of jerk, straub tail, onset of clonus and extensor phases in a dose dependent manner. 300 mg/kg of BMME with 45 mg/kg Pentobarbital showed the highest activity in this test. Isolated compound from BMME also produced significant effect in PTZ induced convulsant model. These findings suggested that the methanol extract of the plant and its isolated bioactive fraction are beneficial in the treatment of epilepsy.

**Keywords:** *Butea monosperma*; Antiepileptic activity; Maximal electroshock; Pentylenetetrazole; Pentobarbital

## Introduction

Traditional, alternative or herbal medicine refers to medical/medication practices other than orthodox medicine/medical practice [1]. The WHO has reported that about 80% of people in the third world countries depend wholly or partly on herbal medicines for their general medications [2]. Many researchers have tried to establish a scientific link/basis to explain the traditional claims of efficacy of most herbs by using different *in vitro* and *in vivo* experimental models. Neurological disorders such as Epilepsy, Parkinson disease, Alzheimer disease, anxiety, depression and other psychiatric disorders have been studied extensively [3-5]. The present study aims to investigate the anticonvulsant activity of methanol stem extract of *Butea monosperma* (Lam.) Kuntze (BMME) and its isoalted bioactive compound by using the Maximal electroshock (MES) convulsion and the Pentylenetetrazole (PTZ) induced convulsion models [6].

The plant *Butea monosperma* (Lam.) Kuntze belongs to the family Fabaceae. The leaves are believed to have astringent, depurative, diuretic and aphrodisiac properties. It promotes diuresis and menstrual flow. The seed is having anthelmintic property. The bark is also used in snakebite as antivenom. Its seeds are when pounded with lemon juice and applied to the skin, it acts as a rubefacient [7]. The main constituents of this plant are butrin, butein, butin, Terpinoids such as lupeol and lupenone. *Butea monosperma* (Lam.) Kuntze also contains flavonoids and steroids. Flowers contain coreopsin, isocoreopsin, sulphurein, monospermoside and isomonospermoside were also identified [8].

Epilepsy is a neurological disorder characterized by recurrent convulsions of cerebral origin, with episodes of sensory, motor or autonomic phenomenon with or without loss of consciousness. About

10% of the patients are refractory and continue to have convulsions at interval of one month or less, which severely disrupts their life and work [9]. Epilepsy is the third most common neurological disorder in the US after Alzheimer's disease and Stroke. Its prevalence is greater than Cerebral palsy, Multiple sclerosis and Parkinson's disease combined [10]. Antiepileptic drugs (AEDs) currently in use notably Phenytoin, Carbamazepine, Phenobarbital and Valproate, referred to as older AEDs were (drugs introduced before 1990). These drugs induce hepatic microsomal enzymes, cytochrome P450 (CYPs) thereby complicating the use of multiple anti convulsion drugs as well as impacting metabolism of oral contraceptives, warfarin and many other drugs. The drugs also enhance metabolism of endogenous compounds including gonadal steroids and vitamin D [11]. These shortcomings has led to a renewed reawakening of interest in the search for medicinal plants and their active component that will be effective for the management of diseases including epilepsy.

## Materials and Methods

### Plant collection

Stem of *Butea monosperma* (Lam.) Kuntze was collected from the forest area of Midnapur district, West Bengal, India. The parts were shade dried and herbarium was prepared for the authentication. The powdered stems of *Butea monosperma* (Lam.) Kuntze was authenticated from National Bureau of Plant and Genetic Resources (NBPGR) (Ref. No. NHCP/NBPGR/2010/17), New Delhi. Further, phytochemical investigations was undertaken to know the presence of phytoconstituent in the raw material of *Butea monosperma* (Lam.) Kuntze.

### Extraction

Dried stem powder of *Butea monosperma* (Lam.) Kuntze (70 g)

was exhaustively extracted with 500 mL petroleum ether and then with methanol in Soxhlet apparatus for 48 h. A dark brown residue (9.8 g) was obtained after evaporation of the solvent. The stem extract of *Butea monosperma* (Lam.) Kuntze was used for the further studies.

**Animals** Swiss Albino mice of either sex weighing 25–35 g, 3–4 months of age, were used for the experiment. Animals were housed in group of 6 per cage and maintained in the animal house. They were obtained from Central animal house of Noida Institute of Engineering and Technology (NIET), Greater Noida, Uttar Pradesh. They were given *ad libitum* water and standard pellet diet under strict hygienic conditions. Animals were acclimatized to laboratory conditions before testing e.g., room temperature of  $25 \pm 1^\circ\text{C}$ , relative humidity 45–55% and a 12:12 h light dark cycle.

The experimental protocol was approved from the Institutional Animal Ethics Committee (IAEC) and protocol approval No. is NIET/IAEC/2009/06. The Committee for Purpose of Control and Supervision of Experiments on Animals (CPCSEA) Reg. No. is 1121/ac/07/CPCSEA. All the experimental procedures were carried out according to the proper guidelines of CPCSEA.

**Drugs/chemicals and treatment** Pentobarbitone (Sigma Aldrich, Bangalore), Pure Diazepam was obtained as gift sample from Cipla, Mumbai, Pentylenetetrazole (PTZ) and Carboxy Methyl Cellulose (CMC) were obtained from Sigma Aldrich, Bangalore and CDH, Mumbai respectively. All other chemicals used were of analytical grade.

### Phytochemical screening

Crude methanol extract of *Butea monosperma* was screened for the presence of secondary metabolites such as alkaloids, tannins, saponins, glycosides, flavonoids, steroids and triterpenoids. The screening was done using standard protocol described by Trease and Evans [12,13].

### Acute toxicity studies

The acute oral toxicity study was carried out according to OECD guidelines 423 [14]. First one animal is dosed with 100 mg/kg body weight. If animal dies a much lower dose is tested. If animal survives, then two more animals are dosed, after 48 hrs observation of the first animal. If survives, then the main test should be terminated. If animal dies, two more animals are dosed and observed. The study was performed with an initial dose of 100 mg/kg body weight. The dose sequence followed were 100, 200, 500, 1000 and 2000 mg/kg body weights.

### Pentobarbital-induced sleep in mice

Swiss albino mice of either sex were divided into five groups of six mice each. Group I served as vehicle control, group II served as standard group, group III, IV and V served as test groups (BMME was given in 100, 200 and 300 mg/kg p.o.). Thirty minutes after treatment all the groups were administered with pentobarbital 45 mg/kg i.p. The onset and duration of sleep was observed and recorded with the mice placed in individual cages. Loss of rightening reflex was considered as the criterion for sleep [15] while the interval between the loss and the recovery of straightening was regarded as the duration of sleep [16].

### Pentylenetetrazole (PTZ) induced convulsion in mice (Chemically-induced convulsion model)

Convulsion was induced in mice by administration of PTZ (80 mg/kg i.p.). Jerky movement, straub tail, clonus, extensor were recorded. The animals were divided into 5 groups of 6 animals each [17].

### Maximal electroshock convolution (MES) in mice

Convulsion was induced in the animals by electroconvulsive shock (50 mA for 0.2 s) via a pinna electrode using an electroconvulsometer. The duration of hind limb tonic extension (HLTE) was compared with animals of control group. Decrease in the duration of hind limb extension was considered a protective action. Animals shown convolution and mortality were also recorded. Less convolution and mortality indicated positive action. The animals were divided into 5 groups of 6 animals each. Group I served as control (vehicle) group (treated with 1% w/v carboxy methyl cellulose, 0.25 ml, p.o.). Group II was the reference group and was treated with diazepam (4 mg/kg i.p. 30 min) prior to the induction of convolution. Groups III, IV and V served as test groups and were treated with the BMME (100, 200 and 300 mg/kg p.o. respectively) for 7 days [17].

### Isolation of compound from the methanol extract of stem of *Butea monosperma* (Lam.) Kuntze

The stem extract of *Butea monosperma* (Lam.) Kuntze (100 g) was dissolved in methanol and centrifuged at 2000 rpm for 30 min. The residue was placed on top of the silica gel column (60–120 mesh) and eluted by column chromatography with chloroform : methanol (95:05 v/v), it was further re-chromatographed on silica gel (100–200 mesh) column chromatography resolved into a pure compound eluted by chloroform: methanol (50:50, v/v) [18].

### Statistical analysis

Results were expressed as mean  $\pm$  standard error of mean. Statistical analysis was performed by analysis of variance (ANOVA); when a statistically significant result was obtained with ANOVA, a Dunnets t-test was performed for multiple comparisons. Values of  $p < 0.05$  were considered significant.

## Results

### Acute toxicity studies

According to the OECD guidelines 423, the animals were observed individually after respective dosing, at least once during first 30 min periodically during the first 24 h, and thereafter, for 2 days. Observation includes changes in fur, skin, eye, respiratory, circulatory, central nervous System, mucous membrane, motor activity, behavioural changes, convulsions, salivation, tremors, sleep and mortality were observed. The treated animals did not show any symptoms of toxicity. Hence, BMME was considered to be safe upto 2 g/kg bw. Based upon these, three doses of BMME were selected i.e., high dose (300 mg/kg; bw), medium dose (200 mg/kg; bw) and low dose (100 mg/kg; bw) for *in vivo* pharmacological experiments (Tables 1–4) (Figures 1–3).

### Isolation and characterization

**Isolation of bioactive molecule from the methanol stem extract of stem of *Butea monosperma* (Lam.) Kuntze:** The isolated compound was obtained from the methanol extract of stem of *Butea monosperma* (Lam.) Kuntze as white crystals. The name had given to it was TBM.

### Spectral analysis

**FTIR finger print analysis of isolated compound from the methanol stem extract of *Butea monosperma* (Lam.) Kuntze:** The FTIR spectra was interpreted as follows IR (KBr): 3384.58  $\text{cm}^{-1}$  (Hydrogen bonded OH Stretch), 2942.36  $\text{cm}^{-1}$  (C–H Stretch in  $\text{CH}_2$ ), 1645.85  $\text{cm}^{-1}$  (C=C Symmetric Stretch), 1485.69 (C=C Asymmetric stretch), 1463.19  $\text{cm}^{-1}$  (C–H deformation in  $\text{CH}_2$  and  $\text{CH}_3$ ), 1388.10

S No	Plant constituents	Methanol Extract
1	Saponin	-
2	Carbohydrate	+
3	Tannin	+
4	Flavonoid	+
5	Steroid	++
6	Terpenoid	+++
7	Cardiac Glycoside	-
8	Amino acid	-

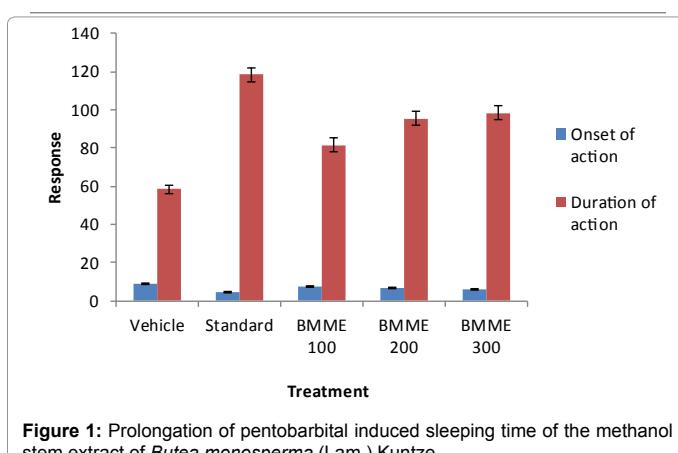
+++ : Significantly Present ; ++: Moderately Present ; +: Slightly Present ; -: Absent

**Table 1:** Preliminary phytochemical studies of methanol extract of stem of *Butea monosperma* (Lam.) Kuntze.

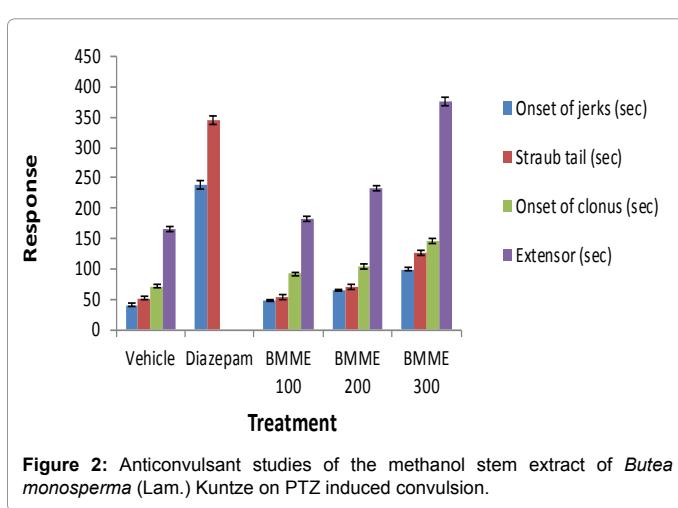
Treatment	Dose (mg/kg)	Onset of action (min)	Duration of action (min)
Vehicle	1 ml/kg	8.77 ± 0.340	58.46 ± 2.213
Chlorpromazine	3	4.84 ± 0.326***	118.40 ± 3.703***
BMME	100	7.86 ± 0.408 <sup>ns</sup>	81.74 ± 3.397***
BMME	200	6.70 ± 0.461**	95.60 ± 3.854***
BMME	300	5.80 ± 0.328***	98.45 ± 3.488***

Values are expressed in Mean ± SEM; \*\*\*: p<0.001, \*\*: p<0.01, ns: not significant, when compared to control group (one way ANOVA followed by Dunnett's 't' test), n=6, BMME represent the methanol extract of the stem of *Butea monosperma* (Lam.) Kuntze.

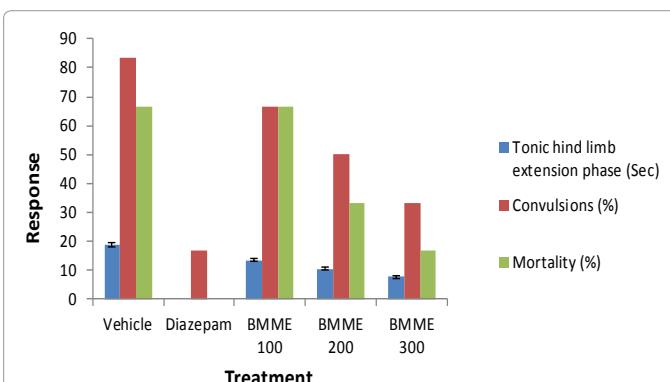
**Table 2:** Prolongation of pentobarbital induced sleeping time of the methanol stem extract of *Butea monosperma* (Lam.) Kuntze.



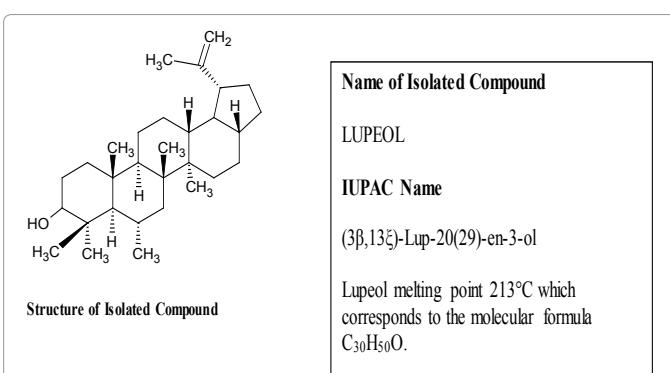
**Figure 1:** Prolongation of pentobarbital induced sleeping time of the methanol stem extract of *Butea monosperma* (Lam.) Kuntze.



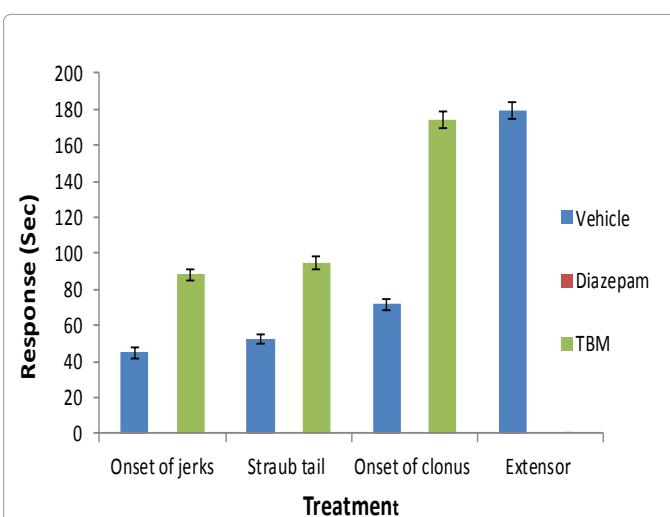
**Figure 2:** Anticonvulsant studies of the methanol stem extract of *Butea monosperma* (Lam.) Kuntze on PTZ induced convulsion.



**Figure 3:** Anticonvulsant studies of the methanol stem extract of *Butea monosperma* (Lam.) Kuntze on MES induced convulsion.



**Figure 4:** Structure and IUPAC name of isolated compound from the methanol stem extract of *Butea monosperma* (Lam.) Kuntze.



**Figure 5:** Anticonvulsant studies of the isolated compound (TBM) from the methanol extract of *Butea monosperma* (Lam.) Kuntze on PTZ induced convulsion.

cm<sup>-1</sup>(C-H Stretch), 1027.79 cm<sup>-1</sup> (C-O Stretch of secondary alcohol), 882.87 cm<sup>-1</sup>(=C-H bending exocyclic CH<sub>2</sub>).

#### NMR spectral study of isolated compound from the methanol stem extract of *Butea monosperma* (Lam.) Kuntze

a) <sup>1</sup>H NMR peak of isolated compound from the methanol stem

**extract of *Butea monosperma* (Lam.) Kuntze:** The <sup>1</sup>H NMR: 7.260 (CDCl<sub>3</sub> peak), 4.683, 4.565(H-29, d, d, 2H), 3.202-3.194 (H, 3, d, d, 1H, 6 Hz, 5 Hz), 2.392(H-19, m, 1H), 2.381 (H-21a, m, 1H), 2.360 (H-15A, t, 1H), 2.348 (H-30, s, 3H), 1.609 (H-12A, 1A, d, 2H), 1.458 (H-13, t, 1H), 1.322 (H-2A, d, 1H), 1.232 (H-2B, q, 1H), 1.198 (H-12A, q, 1H), 1.176 (H-23, s, 3H), 1.080 (H-15A, d, 1H), 0.966 (H-23, s, 3H), 0.942 (H-27, s, 3H), 0.905 (H-18, t, 6 Hz, 1H), 0.787 (H-28, s, 3H), 0.759 (H-24, s, 3H), 0.690 (H-25, s, 3H), 0.671 (H-5, d, 1H).

**b) <sup>13</sup>CNMR peak of isolated compound from the methanol stem extract of *Butea monosperma* (Lam.) Kuntze:** In the <sup>13</sup>C NMR spectrum of Lupeol showed δC: δ 37.354 (C-1), δ 21.108 (C-2), δ 79.178 (C-3), δ 38.241 (C-4), δ 55.481 (C-5), δ 18.171 (C-6), δ 27.626 (C-7), δ 38.886 (C-8), δ 50.625 (C-9), δ 34.463 (C-10), δ 19.476 (C-11), δ 21.108 (C-12), δ 35.760 (C-13), δ 39.031 (C-14), δ 25.334 (C-15), δ 29.857 (C-16), δ 40.176 (C-17), δ 48.493 (C-18), δ 48.159 (C-19), δ 151.135 (C-20), δ 27.596 (C-21), δ 38.886 (C-22), δ 25.334 (C-23), δ 15.523 (C-24), δ 16.145 (C-25), δ 15.98 (C-26), δ 14.719 (C-27), δ 16.282 (C-28), δ 109.477 (C-29) and δ 18.490 (C-30).

**Mass spectra of isolated compound from the methanol stem extract of *Butea monosperma* (Lam.) Kuntze:** The bioactive isolated compound from BAME had given molecular ion peak [M+1] 425.379 in ESI mass spectrum suggested molecular formula C<sub>30</sub>H<sub>50</sub>O.

On the basis of phytochemical screening the isolated compound was found to be a Triterpene. The spectral data of FTIR, NMR and MASS were interpreted to predict the molecular structure, atomic stretching, possible molecular functional groups, etc. The compound thus interpreted was lupeol, a triterpenoid, isolated compound from stem of *Butea monosperma* (Lam.) Kuntze (Figures 4 and 5).

The isolated compound, TBM from BMME was screened to

establish its effect on PTZ induced convulsion. In PTZ induced convulsions onset time of jerk, straub tail, clonus and extensor were significantly increased. The results were better enough than the effects given by BMME on the respective parameters (Table 5).

## Discussion

Phytochemical screening of the extract revealed the presence of secondary metabolites such as terpenoids, tannins, glycosides and flavonoids. Anticonvulsant effect of saponins and flavonoids has been reported [19,20]. The sub acute toxicity study showed no sign and symptoms of toxicity. The extract dose dependently reduced the onset of pentobarbital induced sleep, pentobarbital acts by increasing GABA mediated synaptic inhibition either by directly activating GABA receptors or, more usually, by enhancing the action of GABA on GABA<sub>A</sub> receptors [9]. The total sleep time was potentiated significantly at the dose 300 mg/kg suggesting that the extract of *Butea monosperma* has sleep potentiating properties [21].

*Butea monosperma* extract produced 50-100% protection of the mice against PTZ induced convolution at doses of 100-300 mg/kg. The protection of the extract against PTZ induced convolution suggested that the extract interacts with GABA-ergic neurotransmission. The PTZ test is assumed to identify anticonvulsant drugs effective against myoclonic and absence convulsions. *Butea monosperma* significantly attenuated electrically induced convolution in mice. Electroshock convulsions are characterized by tonic extension of the hind limb and abolition of this activity is taken as anticonvulsant action. The ability of *Butea monosperma* to decrease the duration of tonic clonic convolution in the MES test shows its activity against generalized tonic clonic convulsions [22-25]. *Butea monosperma* has demonstrated potent activity against PTZ and MES convulsions and it would generally be right to say that

Treatment	Dose (mg/kg)	Onset of jerks (sec)	Straub tail (sec)	Onset of clonus (sec)	Extensor (sec)
Control	Vehicle	40.67 ± 1.978	51.17 ± 3.167	70.83 ± 3.060	165.50 ± 3.998
Diazepam	4	238.16 ± 7.234***	344.80 ± 7.317**	No	No
BMME	100	48.00 ± 1.880*	53.67 ± 3.073 <sup>ns</sup>	91.50 ± 3.394***	181.80 ± 4.826*
BMME	200	64.50 ± 1.875**	70.00 ± 3.406***	104.50 ± 3.992***	232.80 ± 4.159***
BMME	300	99.67 ± 2.813***	125.50 ± 3.742***	146.5 ± 4.530***	375.00 ± 7.147***

Values are expressed in Mean ± SEM; \*\*\*: p<0.001, \*\*: p<0.01, \*: p<0.05, ns: not significant, when compared to control group (one way ANOVA followed by Dunnet's 't' test), n=6, BMME represent the methanol extract of the stem of *Butea monosperma* (Lam.) Kuntze.

**Table 3:** Anticonvulsant studies of the methanol stem extract of *Butea monosperma* (Lam.) Kuntze on PTZ induced convolution.

Treatment	Dose (mg/kg p.o.)	Tonic hind limb extension phase (Sec)	Convulsions (%)	Mortality (%)
Vehicle Control	Vehicle	18.83 ± 0.803	83.33	66.67
Diazepam	4	No	16.67	No
BMME	100	13.48 ± 0.446***	66.67	66.67
BMME	200	10.52 ± 0.572***	50.00	33.33
BMME	300	07.733 ± 0.466***	33.33	16.67

Values are expressed in Mean ± SEM; \*\*\*: p<0.001, \*\*: p<0.01, ns: not significant, when compared to control group (one way ANOVA followed by Dunnet's 't' test), n=6, BMME represent the methanol extract of the stem of *Butea monosperma* (Lam.) Kuntze.

**Table 4:** Anticonvulsant studies of the methanol stem extract of *Butea monosperma* (Lam.) Kuntze on MES induced convolution.

Treatment	Dose (mg/kg)	Onset of jerks (sec)	Straubtail (sec)	Onset of clonus (sec)	Extensor (sec)
Control	Vehicle	44.83 ± 3.146	52.67 ± 2.459	71.83 ± 3.229	179.20 ± 4.126
Diazepam	4	No	No	No	No
Isolated compound	20	88.17 ± 3.371***	94.50 ± 3.793***	174.00 ± 4.865***	No

Values are expressed in Mean ± SEM; \*\*\*: p<0.001, when compared to control group (one way ANOVA followed by Dunnet's 't' test), n=6, isolated compound obtained from the methanol extract of the stem of *Butea monosperma* (Lam.) Kuntze.

**Table 5:** Anticonvulsant studies of the isolated compound (TBM) from the stem of *Butea monosperma* (Lam.) Kuntze on PTZ induced convolution.

it will be effective against absence convulsions and generalized tonic-clonic convulsions. Since the MES test identifies agent with activity against generalized tonic-clonic convulsions, whereas PTZ induced convolution model identifies agent with activity against myoclonic and generalized seizure by evaluating onset of jerk, straub tail, onset of clonus and extensor parameters [24]. The bioactive isolated compound from BMME was confirmed as lupeol by spectral data analysis and possessed anticonvulsant effect in PTZ induced convulsions at 20 mg/kg p.o. may be through GABA<sub>A</sub> mediated action. This indicated that the activity of BMME against convolution was because of triterpene (lupeol) present in the stem of *Butea monosperma* (Lam.) Kuntze.

## Conclusion

In conclusion the result of the present study indicated that the methanol extract of stem of *Butea monosperma* and its isolated bioactive component may be of benefit in the treatment of convolution. These findings further provides validity for the use of the plant for the management of convolution in traditional medicine. Further research is encouraged in the area of determining the mechanism of action of the isolated compound lupeol by biogenic amine levels in mice brain responsible for the antiepileptic activity.

## References

1. Sofowora A (1993) Medicinal plants and traditional medicines in Africa. Chichester John Wiley and Sons, New York. pp: 97-145.
2. World Health Organisation (1996) WHO Guideline for the Assessment of herbal medicines. WHO Expert Committee on specification for pharmaceutical preparation. Technical Report series No. 863, Geneva.
3. Danjuma NM, Abdu-Aguye I, Anuka JA, Hussaini IM, Zezi AU (2009) Evaluation of Anticonvulsant activity of the hydroalcoholic stem bark extract of *Randia nilotica* stapf in mice and chicks. Nig J of Pharm Sci 8: 36-45.
4. Rao RV, Descamps O, John V, Bredsen DE (2012) Ayurvedic medicinal plants for Alzheimers disease. A review Alzheimers Res Ther 4: 22.
5. Auddy Auddy B, Ferreira M, Blasina F, Lafon L, Arredondo F, et al. (2003) Screening of antioxidant activity of three Indian medicinal plants, traditionally used for the anagement of neurodegenerative diseases. J Ethnopharmacol 84: 131-138.
6. Rogawski MA (1992) The NMDA receptor, NMDA antagonists and epilepsy therapy. A status report. Drugs 44: 279-292.
7. Kirtikar KR, Basu BD (1935) Indian medicinal plants. 2nd edn. Lalit Mohan Basu, Allahabad, India 1: 785-788.
8. Gupta SR, Ravindranath B, Seshadri T (1970) The glucosides of *Butea monosperma*. Phytochemistry 9: 2231-2235.
9. Rang HP, Dale MM, Ritter JM, Flower RJ, Moore PK (2011) Antiepileptic drugs. In Pharmacology. 7th edn. Churchill Livingstone, London. pp: 540-552.
10. Epilepsy Foundation about Epilepsy (2011).
11. Kwan P, Brodie MJ (2000) Early identification of refractory epilepsy. NEJM 342: 314-319.
12. Silva GL, Lee I, Kinghorn AD (1998) Special problems with the extraction of plants. In Cannell RJP ed, Methods in Biotechnology (Natural product Isolation). Humana Press, New Jersey, USA. pp: 245-364.
13. Trease GE, Evans M (1983) Textbook of Pharmacology. 12th edn. Balliere Tindall, London. pp: 322-383.
14. OECD (Organization for Economic Cooperation and Development) Guideline No. 423.
15. Rolland A, Fleurentain J, Lanheres M, Younous C, Misslin R, et al. (1991) Behavioural effects of American traditional plant *Eschscholzia California*: Sedative and anxiolytic properties. Planta medica 57: 212-216.
16. Fujimori H (1965) Potentiation of barbital hypnosis as an evaluation method of central nervous system depressant. Psychopharmacol 7: 374-378.
17. Vogel HG (2002) Drug discovery and evaluation pharmacological assays. 2nd edn. Springer-Verlag, Berlin, Hiedelberg.
18. Gomes A, Chatterjee I, Chakravarty AK (2006) *Daboia russelii* and *Naja kaouthia* venom neutralization by lupeol acetate isolated from the root extract of Indian sarsaparilla *Hemidesmus indicus* R. Br J of Ethnopharmacology 106: 38-43.
19. Shibata S (2001) Chemistry and Cancer preventing Activities of Ginseng saponins and some related triterpenoid compounds. Journal of Korean medical sciences 16: S28-37.
20. Kavvadias D, Sand P, Youdim KA, Qaiser MZ, Rice-Evans C, et al. (2004) The flavones hispidulin, a benzodiazepine receptor ligand with positive allosteric properties traverses the blood brain barrier and exhibit anticonvulsant effects. British Journal of Pharmacology 142: 811-820.
21. Rakotonirina SV, Ngo bum E, Rakotonirina A, Bopelet M (2001) Sedative properties of the decoction of the rhizomes of *Cyperus articulatus*. Fitoterapia 72: 22-29.
22. Loscher W, Schmidt D (1988) Which animal model should be used in the search for new antiepileptic drugs? A proposal based on experimental and clinical considerations. Epilepsy Research 2: 145-181.
23. De Deyn PP, D'Hoope R, Marescau B, Pei YQ (1992) Chemical model for epilepsy with some references to their applicability in the development of anticonvulsants. Epilepsy Research 12: 87-110.
24. White SH, Woodhead JH, Franklin MR, Swinyard EA, Wolf HH (1995) Experimental selection qualification and evaluation of Antiepileptic Drugs. 4th edn. Raven Press, New York. pp: 99-100.
25. Kupferberg HJ, Schmutz M (1998) Screening of new compounds and the role of the pharmaceutical industry. In Engel J, Pedley TA eds., Epilepsy: A Comprehensive Textbook Lippincott-Raven Philadelphia, New York. pp: 1417-1434.